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(54) Title: CALCINEURIN INTERACTING PROTEIN COMPOSITIONS AND METHODS

(57) Abstract

identification Aπ and characterization of a calcineurin interacting (CNI) protein effective enhance immunosuppressive effects of calcineurin-targeted immunosuppressants by potentiating an interaction of an immunophilin with calcineurin is described herein. One embodiment of the invention is the CNI polypeptide encoded by the CNI gene of Saccharomyces cerevisiae. Polynucleotides encoding a CNI protein are also described. Also described are yeast cells carrying mutations in the CNI gene. Further, a method of identifying a small molecule immunosuppressant compound is described. The methods include the use of a cell-based two hybrid protein-protein interaction assay, wherein one of two fusion hybrid proteins in a cell contains a subunit of calcineurin, and the other

GAL -4_{AD} CNIC

0.44kb

1.22 kb

ATG

CNI

Becorv

CNI

3.5 kb

Stop

of two fusion hybrid proteins contains an immunophilin.

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CALCINEURIN INTERACTING PROTEIN COMPOSITIONS AND METHODS

Field of the Invention

The present invention relates to compounds affecting the function of calcineurin, particularly interactions of calcineurin with immunosuppressant drugs.

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30 Background of the Invention

The immune system functions as the body's major defense against diseases caused by invading organisms. This complex system fights disease by killing invaders such as bacteria, viruses, parasites or cancerous cells while leaving the body's normal tissues unharmed. The immune system's ability to distinguish the body's normal tissues, or self, from foreign or

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cancerous tissue, or non-self, is an essential feature of normal immune system function. A second essential feature is memory, the ability to remember a particular foreign invader and to mount an enhanced defensive response when the previously encountered invader returns. The loss of recognition of a particular tissue as self and the subsequent immune response directed against that tissue produce serious illness.

An autoimmune disease results from the immune system attacking the body's own organs or tissues, producing a clinical condition associated with the destruction of that tissue. An autoimmune attack directed against the joint lining tissue results in rheumatoid arthritis; an attack against the conducting fibers of the nervous system results in multiple sclerosis. The autoimmune diseases most likely share a common pathogenesis and the need for safe and effective therapy. One type of therapy that has been employed in combating autoimmune disease is treatment with immunosuppressant drugs, such as cyclosporin A, FK506 and rapamycin. While the treatments are often effective, the drugs typically have undesirable side effects, including neurotoxicity, nephrotoxicity, hypertension, and metabolic disorder. Many of these side effects are due to the drugs' action on cells other than those of the immune system.

In addition to their use in treating autoimmune conditions, immunosuppressive agents have also been used in treating or preventing transplantation rejection. Organ transplantation involving human organ donors and human recipients (allogeneic grafts), and non-human primate donors and human recipients (xenogeneic grafts), has received considerable medical and scientific attention (e.g., Roberts, 1989; Platt, 1990). To a great extent, this effort has been aimed at eliminating, or at least reducing, the problem of rejection of the transplanted organ. In the absence of adequate immunosuppressive therapy, the transplanted organ is destroyed by the host immune system.

Presently, the most commonly used agents for preventing transplant rejection include corticosteroids, cytotoxic drugs that specifically inhibit T cell activation such as azathioprine, immunosuppressive drugs such as cyclosporin A, and specific antibodies directed against T lymphocytes or surface receptors that mediate their activation (Briggs, 1991; Kennedy, 1983; Storb, 1985; Storb, 1986). All of these drug therapies are limited in effectiveness, in part because the doses needed for effective treatment of transplant rejection may increase the patient's susceptibility to infection by a variety of opportunistic invaders, and in part because of direct toxicity and other side effects.

Cyclosporin A, currently the most effective and most commonly used agent, is significantly toxic to the kidney. This nephrotoxicity limits the quantity of drug that can be

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safely given. The physician is frequently forced to administer sub-optimal doses of the drug because of this toxicity. A preparation capable of potentiating the action of immunosuppressive agents such as cyclosporin A on the immune system, thus allowing the administration of lower doses of drug, would be of considerable value in reducing the morbidity and mortality associated with transplantation.

Summary of the Invention

In one embodiment, the present invention includes polypeptide compositions effective to enhance immunosuppressive effects of calcineurin-targeted immunosuppressants by potentiating an interaction of an immunophilin with calcineurin. The present invention includes the isolation and characterization of a calcineurin interacting protein, CNI, having these properties. Also disclosed herein are methods for the isolation and characterization of further CNI-related sequences and sequences of CNI-variants. The amino acid sequences presented as SEQ ID NO:2 and SEQ ID NO:5 are exemplary of the polypeptides of the present invention.

The present invention also includes a CNI polypeptide fragment that interacts specifically with the "A" subunits of calcineurin (CNA1 and CNA2), but not with calcineurin "B" subunit (CNB1). In one embodiment, this fragment has an amino acid sequence of between 15 and 915 amino acids in length, for example, the c-terminal 306 amino acids of the CNI protein (CNIc).

Included aspects of the invention are an CNI polypeptide; a recombinant CNI polypeptide; and a fusion polypeptide comprised of an CNI polypeptide. Exemplary fusion proteins include fusions to β -galactosidase.

The invention further includes isolated nucleic acid sequences encoding the above described polypeptides and polypeptide fragments. Exemplary nucleic acid sequences include the sequences presented as SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:4 and SEQ ID NO:6. The present invention includes CNI-encoding genomic polynucleotides, cDNAs thereto and complements thereof. With respect to polynucleotides, some aspects of the invention include: a purified CNI-encoding genomic polynucleotide; CNI polypeptide-encoding RNA and DNA polynucleotides; recombinant CNI polypeptide-encoding polynucleotides; a recombinant vector including any of the above recombinant polynucleotides, and a host cell transformed with any of these vectors. Another aspect of the invention is a polynucleotide probe for CNI polypeptide-encoding sequences.

Portions of a CNI-polypeptide coding sequences are effective as probes to isolate variants coding sequences which occur naturally, or to determine the presence of such coding

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sequences in nucleic acid samples. Such probes include hybridization screening probes and polymerase chain reaction amplification primers specific for CNI-polypeptide coding sequences. Homologues of CNI may be isolated from a number of sources, such as other types of yeast cells (e.g., Schizosaccharomyces) or mammalian cells (e.g., human).

Other aspects of the invention include: a recombinant expression system which incorporates an open reading frame (ORF) derived from CNI polypeptide-encoding sequences, wherein the ORF is linked operably to a control sequence which is compatible with a desired host, a cell transformed with the recombinant expression system, and a polypeptide produced by the transformed cell. Typically the expression system includes a vector having (a) a nucleic acid containing an open reading frame that encodes a CNI-polypeptide; and (b) regulatory sequences effective to express said open reading frame in a host cell. The regulatory sequence may include sequences useful for targeting or secretion of the CNI-polypeptide: such as a secretory signal recognized in yeast or bacterial expression systems.

The invention includes a method of recombinantly producing CNI-polypeptides. In the method, a recombinant expression system containing an open reading frame (ORF) having a polynucleotide sequence which encodes a CNI-polypeptide, where the vector is designed to express the ORF in the host, is introduced into suitable host cells. The host is then cultured under conditions resulting in the expression of the ORF sequence. The CNI-polypeptide sequences discussed above are examples of suitable CNI-polypeptides. Numerous vectors and their corresponding hosts are useful in the practice of this method of the invention, including, but not limited, to the vectors described herein for expression in yeast cells, and lambda gt11 phage vector and E. coli cells. Other host cells include insect and mammalian cell expression systems.

The invention also includes purified antibodies that are immunoreactive with a CNI-polypeptide. The antibodies may be polyclonal or monoclonal. Antibodies that are specifically immunoreactive with CNI-polypeptides may be useful for the isolation of CNI-polypeptide homologues from other cell type sources (e.g., mammalian).

The present invention also includes, a method of identifying a small molecule immunosuppressant compound. In the method, a cell-based two hybrid protein-protein interaction assay is constructed where one of two fusion hybrid proteins in the cell contains a subunit of calcineurin, and the other of two fusion hybrid proteins contains an immunophilin. The cell is then contacted with the small molecule being tested. A small molecule is identified as an immunosuppressant if the molecule potentiates an interaction between the two hybrid proteins. In one embodiment, the method is carried out using yeast cells, where one of the two

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fusion hybrid proteins contains a GAL4 activation domain and the other of two fusion hybrid proteins contains a GAL4 binding domain. The method can be carried out where subunits of calcineurin are from any cell source, in particular, yeast or mammalian cells (including human cells). The subunit may, for example, be yeast calcineurin subunit CNA1 or CNA2, or human calcineurin subunit "A". The immunophilin can, for example, be cyclophilins or FK506-binding proteins (e.g., FKBP12) typically from a homologous cell source.

Also included in the present invention is another method of identifying a small molecule immunosuppressant compound. In the method, a cell-based two hybrid protein-protein interaction assay is constructed, wherein one of two fusion hybrid proteins in a cell contains an "A" subunit of calcineurin, and the other of two fusion hybrid proteins contains a CNI 10 polypeptide. The cell preferably, but not necessarily, also contains a vector construct causing overexpression, or increased expression, of a "B" subunit of calcineurin. The cell is then contacted with the small molecule being tested. A small molecule is identified as an immunosuppressant if the molecule potentiates an interaction between the two hybrid proteins. This method is used to identify compounds (like FK506) that potentiate the interaction between 15 CNI and CNA1. In one embodiment, the method is carried out using yeast cells, where one of the two fusion hybrid proteins contains a GAL4 activation domain and the other of two fusion hybrid proteins contains a GAL4 binding domain. The method can be carried out where subunits of calcineurin are from any cell source, in particular, yeast or mammalian cells (including human cells). The subunits may, for example, be calcineurin subunit A1 or A2. 20 The CNI polypeptide may also be from any source (e.g., yeast or human), and may be only a fragment of a complete CNI polypeptide (such as a c-terminal fragment). An exemplary cterminal fragment of CNI is CNIc.

Further, included in the present invention, is a yeast cell carrying a mutation in the naturally-occurring copy of CNI, where the mutation prevents expression of a functional CNI protein from the genomic copy. Embodiments of this aspect of the present invention include deletion mutations within the coding region of the CNI gene, deletion of regulation regions of the CNI gene, and non-sense or mis-sense mutations in the CNI gene. Yeast cells having such mutations are useful, for example, in a method of identifying proteins of similar function to CNI. In one embodiment, a hybrid interaction screen is set up in a cell with a CNI deletion and a GAL4 protein binding domain-CNA fusion and a GAL4 activation domain-immunophilin fusion. Expression libraries are then screened to identify clones encoding proteins that potentiate an interaction of an immunophilin with calcineurin. This screen will identify CNI-coding sequences as well as other proteins with a similar function.

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In a related embodiment, a yeast cell with a CNI deletion is used to identify CNI homologues (e.g., from other organisms, such as human) using a complementation assay or screen. Expression libraries (e.g., human lymphocyte expression libraries) are transformed into cells with a CNI deletion, and transformants are selected on their ability to complement the function of yeast CNI. An exemplary assay for selecting such transformants is exposure to hygromycin B. Cells which become more sensitive to hygromycin B following transformation are further analyzed to determine if the plasmid with which they were transformed contains an insert homologous to yeast CNI, or encoding a polypeptide with similar function to CNI.

The invention also includes a yeast cell carrying a mutation in the naturally-occurring genomic copy of a gene encoding calcineurin-interacting polypeptide, where the mutation prevents expression of a functional calcineurin-interacting polypeptide from the genomic copy. The mutation may be a null mutation, such as described in Example 8 below, or a different type of mutation, e.g., a nonsense or missense mutation. Nonsense and missense mutations may be generated using standard methods.

These and other objects and features of the invention will be more fully appreciated when the following detailed description of the invention is read in conjunction with the accompanying drawings.

Brief Description of the Figures

Figures 1A and 1B present schematic diagrams of sequences encoding the c-terminal portion of CNI (CNIc) fused to GAL-4 activation domain (GAL-4AD) (Fig. 1A), and sequences encoding CNI (Fig. 1B).

Figure 2A presents data from a β -galactosidase (β -gal) assay to detect the interaction of CNIc with the A1 subunit of calcineurin (CNA1), A2 subunit of calcineurin (CNA2), GAL-4 binding domain (G4BD) and lamin C. A labeled schematic diagram corresponding to the data shown in Fig. 2A is presented in Fig. 2B to facilitate reference to individual groups of colonies.

Figure 3A presents data from a β -gal assay to detect the interaction of CNIc with CNA1 Δ C, CNA2 Δ C and CNB1. A labeled schematic diagram corresponding to the data shown in Fig. 3A is presented in Fig. 3B.

Figures 4A and 4C present data from β -gal assays to evaluate the effects of FK506 and the deletion of CNB1 on the interactions of CNIc with CNA and CNA Δ C. Labeled schematic diagrams corresponding to the data shown in Figs. 4A and 4C are presented in Figs. 4B and 4D, respectively.

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Figures 5A and 5C present data from β -gal assays to evaluate the effects of FK506 and the overexpression of CNB1 on the interactions of CNIc with CNA and CNA Δ C. Labeled schematic diagrams corresponding to the data shown in Figs. 5A and 5C are presented in Figs. 5B and 5D, respectively.

Figures 6A, 6C and 6E present data from β -gal assays to evaluate the effects of FK506, cyclosporin A (CsA), and the deletion of CNB1 on the interaction of CNIc with CNA and CNA \triangle C. Labeled schematic diagrams corresponding to the data shown in Figs. 6A, 6C and 6E are presented in Figs. 6B, 6D and 6F, respectively.

Figures 7A, 7C, 7E and 7G present data from β -gal assays to evaluate the effects of FK506, cyclosporin A (CsA), rapamycin and the overexpression of CNB1 on the interaction of CNIc with CNA and CNA Δ C. Labeled schematic diagrams corresponding to the data shown in Figs. 7A, 7C, 7E and 7G are presented in Figs. 7B, 7D, 7F and 7H, respectively.

Figure 8A presents data from a β -gal assay to evaluate the effects of overexpression of full-length CNI on FK506-dependent interaction of FKBP with CNA. A labeled schematic diagram corresponding to the data shown in Fig. 8A is presented in Fig. 8B.

Figures 9A and 9C present data from β -gal assays to evaluate the effects of overexpression of full-length CNI on FK506-dependent interaction of FKBP with CNB1. Labeled schematic diagrams corresponding to the data shown in Figs. 9A and 9C are presented in Figs. 9B and 9D, respectively.

Figure 10 presents an image of a protein blot of CNIc and CNA co-immunoprecipitate probed with anti-CNA2 antibody.

Figure 11 presents an image of a yeast RNA blot hybridized with a CNIc probe.

Figure 12 presents the DNA sequence of a 3.5 kb fragment of yeast chromosome 11 containing the coding sequence for a yeast CNI protein.

Figure 13 presents the amino acid sequence of a yeast CNI protein.

Brief Description of the Sequences

SEQ ID NO:1 presents the nucleotide sequence of a Sau3AI fragment containing the coding sequence for CNIc.

SEQ ID NO:2 presents the amino acid sequence of CNIc encoded by SEQ ID NO:1.

SEQ ID NO:3 presents the coding sequence presented in SEQ ID NO:1.

SEQ ID NO:4 presents the nucleotide sequence of a gene encoding a complete CNI protein.

SEQ ID NO:5 presents the amino acid sequence encoded by SEQ ID NO:4.

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SEQ ID NO:6 presents the coding sequence presented in SEQ ID NO:4.

SEQ ID NO:7 presents the nucleotide sequence of PCR primer CNI-PCR-A.

SEQ ID NO:8 presents the nucleotide sequence of PCR primer CNI-PCR-B.

SEQ ID NO:9 presents the nucleotide sequence of a gene encoding the yeast CNA1 subunit of calcineurin.

SEQ ID NO:10 presents the amino acid sequence encoded by SEQ ID NO:9.

SEQ ID NO:11 presents the nucleotide sequence of a gene encoding the yeast CNA2 subunit of calcineurin.

SEO ID NO:12 presents the amino acid sequence encoded by SEQ ID NO:11.

SEQ ID NO:13 presents the nucleotide sequence of a gene encoding the yeast CNB1 subunit of calcineurin.

SEQ ID NO:14 presents an amino acid sequence encoded by SEQ ID NO:13.

SEQ ID NO:15 presents an amino acid sequence encoded by SEQ ID NO:13.

SEQ ID NO:16 presents the coding sequence presented in SEQ ID NO:13.

SEQ ID NO:17 presents the amino acid sequence encoded by SEQ ID NO:16.

SEO ID NO:18 presents a nucleotide sequence encoding CNA1Δc.

SEQ ID NO:19 presents the amino acid sequence encoded by SEQ ID NO:18.

SEQ ID NO:20 presents a nucleotide sequence encoding CNA2Δc.

SEQ ID NO:21 presents the amino acid sequence encoded by SEQ ID NO:20.

SEQ ID NO:22 presents the nucleotide sequence of PCR primer G4-PCR-A.

SEQ ID NO:23 presents the nucleotide sequence of PCR primer G4-PCR-B.

Detailed Description of the Invention

I. **DEFINITIONS**

A "calcineurin-targeted immunosuppressant" is a compound that possesses in vivo immunosuppressive activity, and that interacts with an immunophilin to form a complex which is capable of inhibiting calcineurin.

"Interacting proteins" are proteins capable of specifically binding to one another, or associating with one another, in a cell or *in vitro*.

A calcineurin interacting (CNI) protein or polypeptide is a protein or polypeptide that is effective to enhance immunosuppressive effects of a calcineurin-targeted immunosuppressant by potentiating an interaction of an immunophilin with calcineurin. Preferably, a CNI protein or polypeptide is a protein or polypeptide having an amino acid sequence that is homologous to the sequence presented herein as SEQ ID NO:5.

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"Substantially isolated" is used in several contexts and typically refers to the at least partial purification of a CNI protein or polypeptide fragment away from unrelated or contaminating components (e.g., cytoplasmic contaminants and heterologous proteins). Methods and procedures for the isolation or purification of compounds or components of interest are described below (e.g., affinity purification of fusion proteins and recombinant production of CNI polypeptides).

In the context of the present invention, the phrase "nucleic acid sequences," when referring to sequences which encode a protein, polypeptide, or peptide, is meant to include degenerative nucleic acid sequences which encode homologous protein, polypeptide or peptide sequences as well as the disclosed sequence.

Two nucleic acid fragments are considered to have "homologous" sequences if they are capable of hybridizing to one another (i) under typical hybridization and wash conditions, as described, for example, in Sambrook, et al., pages 320-328, and 382-389, or (ii) using reduced stringency wash conditions that allow at most about 25-30% basepair mismatches, for example: 2 × SSC, 0.1% SDS, room temperature twice, 30 minutes each; then 2 × SSC, 0.1% SDS, 37°C once, 30 minutes; then 2 × SSC, room temperature twice, 10 minutes each. Preferably, homologous nucleic acid strands contain 15-25% basepair mismatches, even more preferably 5-15% basepair mismatches. These degrees of homology can be selected by using wash conditions of appropriate stringency for identification of clones from gene libraries (or other sources of genetic material), as is well known in the art.

Two amino acid sequences or two nucleotide sequences (in an alternative definition for homology between two nucleotide sequences) are considered homologous (as this term is preferably used in this specification) if they have an alignment score of >5 (in standard deviation units) using the program ALIGN, typically default mutation gap matrix and gap penalty (Dayhoff). The two sequences (or parts thereof) are more preferably homologous if their amino acids are greater than or equal to 40% using the ALIGN program mentioned above.

II. OVERVIEW OF INVENTION

Experiments performed in support of the present invention demonstrate the identification and isolation of the nucleic acid sequence encoding a calcineurin interacting (CNI) protein. Further experiments performed in support of the present invention characterize the CNI protein, as well as a polypeptide containing only the c-terminal 306 amino acids of the CNI protein (CNIc). The experiments demonstrate that CNIc interacts specifically with the "A" subunits of calcineurin (CNA1 and CNA2), but not with calcineurin "B" subunit (CNB1).

The experiments also demonstrate that CNIc does not interact directly with FK506 binding protein (FKBP; Schreiber, et al.; with or without FK506), GAL4 binding domain (G4_{BD}) or lamin C. The experiments also demonstrate that CNIc interacts with C-terminally truncated forms of CNA (CNA Δ C), which have lost their autoinhibitory domains, though the interaction is somewhat weaker that with full length CNA proteins.

Additional experiments show that the interaction between CNIc and CNA is enhanced when CNB1 is deleted, and diminished when CNB1 is overexpressed, that the interaction between CNIc and CNA or CNA Δ C is markedly enhanced by FK506 and by Cyclosporin A (CsA), but not rapamycin, and that overexpression of a full-length CNI protein enhances the interaction between CNA and FKBP (detectable only in the presence of FK506).

Additional experiments conducted in support of the present invention demonstrate that overexpression of the full-length CNI has no detectable effect on the interaction between CNB1 and CNA, and that in the presence of FK506 or CsA, overexpression of CNB1 no longer inhibits the interaction of CNIc with CNA.

It was also found that CNI deletion mutants are viable, both in wild-type and CN-deletion backgrounds, and that CNI deletion mutants in a CN-deletion background are more resistant to hygromycin B than normal CN-deletion mutants.

Co-immunoprecipitation experiments demonstrate that CNIc and CNA co-immunoprecipitate in the presence of FK506, and protein blot experiments show that CNI is expressed at low levels *in vivo*. RNA blot experiments show that CNI is encoded by a single message approximately 2.9 kb in length.

A comparison of the yeast CNI sequence with sequences present in nucleic acid and amino acid databases reveals no obvious homologous sequences have been identified in other organisms.

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III. <u>CALCINEURIN</u>

Experiments performed in support of the present invention were designed to identify polypeptides capable of interacting with calcineurin. Calcineurin (also called phosphoprotein phosphatase 2B or PP2B), has been characterized from many different tissues and organisms (Klee, et al.). It is a heterodimer of two subunits, of which the "A" subunit is about 61 kD in weight, possesses catalytic activity and also contains the association site for calmodulin. The "B" subunit contains four Ca2+ binding sites and activates the A subunit. Calcineurin has little enzymatic activity, even in the presence of Ca2+ and only becomes fully active when associated with calmodulin (Cyert).

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Two A subunits (CNA1 and CNA2; Cyert, et al., 1991) and one B subunit (CNB1, Cyert, et al., 1992) have been cloned in yeast. Either CNA1 or CNA2 may associate with CNB1 to form a functional calcineurin heterodimer. Multiple isotypes of the A subunit have been cloned from a variety of organisms and are highly conserved (Klee, et al.). In particular, calcineurin subunits have been cloned from human tissue (see reviews by, for example, Klee, et al., and Guerini, et al.).

IV. IMMUNOSUPPRESSANT DRUGS

FK506, cyclosporin A (CsA) and rapamycin, derived from fungi, inhibit the activation of T-cells by antigens. The compounds have proven highly effective at suppressing mammalian immune systems in vivo. In particular, CsA therapy in clinical settings has dramatically increased the success rate of transplantation therapy.

It is now known that FK506 and CsA exert their immunosuppressive effects, in part, by inhibiting the transcriptional activation of the interleukin-2 (IL-2) gene, whereas rapamycin appears to function by inhibiting the response of T-cells to IL-2, presumably by inhibiting a transduction pathway mediated by the IL-2 receptor.

The molecular mechanism of FK506 and CsA immunosuppressive action involves a group of small, abundant intracellular proteins termed immunophilins, which bind with a high affinity to the immunosuppressants (Schreiber). At least two classes of immunophilins are known to exist. One class, termed cyclophilins, binds to CsA, while another class, the FK506-binding proteins (FKBPs) binds FK506 and rapamycin. Many immunophilin genes, from a variety of organisms, have been cloned, and appear to be highly conserved from simple eukaryotes to mammals.

It is believed that FK506 and CsA-induced immunosuppression is due to the binding of complexes, formed by binding of immunosuppressants FK506 and CsA bound to one of their respective immunophilins, to the catalytic subunit of calcineurin (Schreiber, et al., Liu, et al., Foor, et al., Weiss, et al.). The binding of such a complex to an (A) subunit inhibits activation of calcineurin by increased intracellular calcium, which in turn prevents calcineurin from activating transcription factor NF-AT. Since IL-2 is one of the genes controlled by NF-AT in T-cells, inhibition of the transcription factor inhibits the production of IL-2, resulting in immunosuppression (Clipstone, et al.).

FK506 and CsA are widely used in organ transplantation to prevent host rejection. However, both drugs are known to have many undesired side-effects such as neurotoxicity, nephrotoxicity, hypertension, and metabolic disorder. Accordingly formulations effective to

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increase a target cell's sensitivity to these drugs may be useful in alleviating some of the aforementioned side-effects. Specifically, CNI and its homologues or derivatives, administered at appropriate levels, may be able to increase the sensitivity of CN to FK506/CsA and reduce the necessary dosage thus reducing or eliminating the side-effects of these drugs.

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V. Two Hybrid Protein Interaction Assays

Two hybrid protein interaction assay methods (two hybrid protein-protein interaction screens) provide a simple and sensitive means to detect the interaction between two proteins in living cells. The assays are based on the finding that most eukaryotic transcription activators are modular (e.g, Brent, et al.), i.e., that the activators typically contain activation domains that activate transcription, and DNA binding domains that localize the activator to the appropriate region of a DNA molecule.

The development of two hybrid protein interaction assays was made possible by the observation that the DNA binding domain does not need to be physically located on the same polypeptide as the activation domain (Ma, et al., Triezenberg, et al.), raising the possibility that transcription of reporter genes could be used as an assay to detect protein interactions.

The utility of two hybrid systems for detecting interactions between two interacting proteins was fully realized by the observation that protein interactions could be detected if two potentially-interacting proteins were expressed as fusions, or chimeras (Fields, et al.). A first fusion protein contains one of a pair of interacting proteins fused to a DNA binding domain, and a second fusion protein contains the other of a pair of interacting proteins fused to a transcription activation domain. The two fusion proteins are independently expressed in the same cell, and interaction between the "interacting protein" portions of the fusions reconstitute the function of the transcription activation factor, which is detected by activation of transcription of a reporter gene.

At least two different cell-based two hybrid protein-protein interaction assay systems have been used to assess binding interactions and/or to identify interacting proteins. Both employ a pair of fusion hybrid proteins, where one of the pair contains a first of two "interacting" proteins fused to a transcription activation domain of a transcription activating factor, and the other of the pair contains a second of two "interacting" proteins fused to a DNA binding domain of a transcription activating factor.

The yeast GAL4 two hybrid system (Fields, et al., Chien, et al., Durfee, et al., Bartel, et al.), utilized for experiments performed in support of the present invention, was developed

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to detect protein-protein interaction based on the reconstitution of function of GAL4, a transcriptional activator from yeast, by activation of a GAL1-lacZ reporter gene.

Like several other transcription activating factors, GAL4 contains two distinct domains, a DNA binding domain and a transcription activation domain. Each domain can be independently expressed as a portion of a fusion protein composed of the domain, and a second, "bait" interacting protein. The two fusion proteins are then independently expressed together in a cell. When the two GAL4 domains are brought together by a binding interaction between the two "interacting" proteins, transcription of a reporter gene under the transcriptional control of GAL4 is initiated. The reporter gene typically has a promoter containing GAL4 protein binding sites (GAL upstream activating sequences, UAS_G). Exemplary reporter genes are the GAL1-lacZ, and GAL1-HIS3 reporter genes used in experiments described herein.

A second two hybrid system, described in detail in Ausubel, et al., utilizes a native E. coli LexA repressor protein, which binds tightly to appropriate operators. A plasmid is used to express one of a pair of interacting proteins (the "bait" protein) as a fusion to LexA. The plasmid expressing the LexA-fused bait protein is used to transform a reporter strain of yeast, such as EGY48, that contains pSH18-34.

In this strain, binding sites for LexA are located upstream of two reporter genes. In the first reporter system, the upstream activation sequences of the chromosomal LEU2 generequired in the biosynthetic pathway for leucine (Leu)—are replaced in EGY48 with lexA operators, permitting selection for viability when cells are plated on medium lacking Leu. In the second reporter system, EGY48 harbors a plasmid, pSH18-34, that contains a lexA operator-lacZ fusion gene, permitting discrimination based on color when the yeast is grown on medium containing Xgal (Ausubel, et al.).

To screen a library with the LexA system, the library uses the inducible yeast GAL1 promoter to express proteins as fusions to an acidic domain ("acid blob") that functions as a portable transcriptional activation motif ("act"), and to other useful moieties. Expression of library-encoded proteins is induced by plating transformants on medium containing galactose (Gal), so yeast cells containing library proteins that do not interact specifically with the bait protein fail to grow in the absence of Leu. Yeast cells containing library proteins that interact with the bait protein form colonies within 2 to 5 days, and the colonies turn blue when the cells are streaked on medium containing Xgal. The plasmids are isolated and characterized by a series of tests to confirm specificity of the interaction with the initial bait protein. Those found to be specific are ready for further analysis (e.g., sequencing).

LexA and GAL4 each have different properties that should be considered when selecting a system. LexA is derived from a heterologous organism, has no known effect on the growth of yeast, possesses no residual transcriptional activity, can be used in GAL4⁺ yeast, and can be used with a Gal-inducible promoter. Because GAL4 is an important yeast transcriptional activator, experiments must be performed in gal4 yeast strains to avoid background from endogenous GAL4 activating the reporter system.

Both two hybrid systems have been successfully used for isolating genes encoding proteins that bind a target protein and as simple protein binding assays (e.g., Yang, et al., Gyuris, et al.), and both can be applied to methods of the present invention.

Both gene isolation and protein binding assay applications of the GAL4 system are described in Examples below.

VI. SPECIFIC EMBODIMENTS

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Example 1 demonstrates application of an exemplary two hybrid protein-protein interaction screen (Materials and Methods, section D) to the screening of three pGAD yeast fusion libraries, carrying fusions between the transcription activating domain of yeast protein GALA (G4AD) and yeast genomic DNA Sau3Al fragments, in all three reading frames. The libraries are screened to identify polypeptides, encoded by the Sau3Al fragments, capable of interacting with catalytic (A) subunits of calcineurin, expressed as fusions with the GALA protein binding domain (GBT-CNA fusions).

Three sets of yeast cells harboring pGBT-CNA1 TRP1 (GBT-A1) hybrid plasmid and a GAL4-activated LacZ reporter gene are each transformed with one of the three reading-frame libraries. Construction of the plasmids used is described in Materials and Methods, sections B and C. Cells transformed with a plasmid encoding a protein fusion capable of interacting with the CNA subunit fusion are selected using a β-galactosidase (β-gal) assay on plates containing the chromogenic substrate X-gal (Materials and Methods, section E). Results of the β-gal assay are confirmed using a growth assay (Materials and Methods, section F). False positives are eliminated by colony purification (re-streaking for single colonies), PCR experiments using GAL4 primers, and testing against a number of test fusions by β-gal assays on transformed haploid or mated diploid reporter strains.

A yeast clone encoding a polypeptide capable of specifically interacting with CNA polypeptide fusions is identified and sequenced. The sequence of the Sau3AI fragment is presented as SEQ ID NO:1. The coding sequence forming the open reading frame is presented as SEQ ID NO:3. The polypeptide encoded by the open reading frame is presented as SEQ

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ID NO:2. The open reading frame encodes 306 amino acids followed by a stop codon in frame with the coding sequence of GAL4, but does not contain an in-frame ATG start site upstream of the stop codon. This is consistent with the fragment encoding a c-terminal portion of a complete gene product. The clone is termed CNIc, with the lowercase "c" representing "c-terminal".

Figure 1A shows a schematic representation of the nucleic acid sequence encoding the GAL4AD-CNIc fusion protein. The stippled portion between GAL4AD and CNIc represents a linker discussed in the Materials and Methods section, as well as in Example 1. Also indicated in the Figure are the approximate locations of in-frame start (ATG) and stop codons, and Sau3AI restriction sites.

Example 1 further describes the identification of a λ clone encoding a full length sequence version of CNIc, termed CNI. The polypeptide encoded by the sequence is termed CNI protein. The clone is identified by hybridization screening of a panel of λ clones spanning the yeast genome using a 1.22 kb ³²P-labeled probe generated from CNIc.

Phage lysates of the λ clone are amplified, purified, restriction-mapped and used as a DNA source for subsequent cloning experiments. A 3.16 kb MunI/EcoRV fragment from the λ clone insert contains the coding sequence of CNI. The sequence of the 3.16 kb MunI/EcoRV fragment is encompassed by the 3.5 kb sequence presented as SEQ ID NO:4, and a schematic diagram of the sequence is shown in Figure 1B. This sequence contains the entire 2.75 kb coding sequence of CNI (SEQ ID NO:6; graphically indicated in Figure 1B by the locations of the "ATG" and "Stop" codons).

A search of known DNA and protein sequences turns up no obvious matches or homologies to genes in other organisms. Accordingly, CNI may represent a new type of calcineurin interacting protein.

The methods referred to above may also be applied to the screening of, for example, a human cDNA library using an appropriate two-hybrid protein interaction screen. The "bait" protein in the interaction screen (e.g., the protein analogous to CNA1 in Example 1) may be of yeast origin (e.g., CNA1), but is preferably of human origin (e.g., a human calcineurin "A" subunit; Klee, et al.). The bait protein is expressed in the cell (e.g., a yeast cell) used for the two hybrid interaction screen as a fusion to a domain of a transcription activating factor (e.g., the DNA binding domain of GAL4). The library may be a human DNA library in a vector (e.g., pGAD) effective to express library sequences as fusions to a complimentary domain of the transcription activating factor (e.g., the activation domain of GAL4). Libraries of human sequences can be derived from a number of sources including genomic DNA, such as yeast

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artificial chromosome (YAC) constructs carrying genomic human DNA, or cDNA generated from a variety of cell types (e.g., activated T-cells).

Example 2 details a β -gal assay to determine the specificity of binding of CNIc to subunits of calcineurin. Exemplary results are shown in Figure 2A. The legend for Fig. 2A is shown in Figure 2B. Numbers in the legend refer to locations of yeast colonies expressing particular combinations of plasmid constructs (indicated in Example 2).

A comparison of the intensities of the blue β -gal reaction product indicates that CNIc interacts strongly with CNA1 (21, 22 and 23), and somewhat less strongly with CNA2 (24). Neither cells containing AS-lamin with GAD-CNIc (25), nor cells containing only GBT with GAD-CNIc (26) show a detectable signal above background. Two subunits of calcineurin (CNA1, CNB1) known to interact with each other are used as a positive control for the assay (20). The data presented in Figure 2A show that CNIc interacts specifically with CNA1 and CNA2, but not with G4BD or lamin C.

A similar set of experiments, illustrated in Fig. 3A, is conducted using constitutively-active CNA subunits, as well as calcineurin subunit CNB1. As described in the Materials and Methods section below, CNA1 Δ C and CNA2 Δ C are each missing a C-terminal portion of the protein containing an autoinhibitory domain. Exemplary results from these experiments are shown in Figure 3A. The legend for Fig. 3A is shown in Figure 3B.

The data show that GBT-A1 Δ C and GAD-CNIc (28) gives a definite positive signal, while GBT-A2 Δ C and GAD-CNIc (29) is weaker, though still detectable above its background (i.e. GBT-A2 Δ C and GAD; 31). The signal from GBT-B1 and GAD-CNIc (32) is not detectable above vector background (GBT-B1 and GAD; 33). Positive controls GBT-A1 Δ C and GAD-B1 (34) and GBT-A2 Δ C and GAD-B1 (35) give strong signals. The data presented in Figure 3A show that CNIc interacts specifically with CNA1 Δ C and CNA2 Δ C, but not with CNB1.

Example 3 details the effects of immunosuppressant drugs on binding of CNIc to calcineurin in $B1^{\infty}$, B1 Deletion and B1 Overproducing Yeast Strains. The yeast strains are assayed for β -gal activity as above to determine if the immunosuppressant drugs FK506, cyclosporin A (CsA) and rapamycin affect the binding of CNIc to subunits of calcineurin. The experiments are performed in yeast strains wild-type for the CNB1 subunit, null for the CNB1 subunit, and in yeast transformed with a high efficiency expression vector containing DNA encoding the CNB1 subunit.

Exemplary data are shown in Figures 4A, 4C, 5A and 5C. The legends for Figs. 4A and 4C are shown in Figures 4B and 4D, respectively. Figs. 4A and 5A illustrate experiment

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performed without FK506, while experiments shown in Figs. 4C and 5C were performed with FK506.

The interactions of various combinations of proteins expressed by constructs indicated in Example 3 was studied in three yeast strains, one of which is null for the CNB1 subunit of calcineurin (Y153b; at 36-40), while the others (Y190; at 41-46 and Y526 at 47-51) are wild-type for CNA1, CNA2 and CNB1.

The data, shown in Fig. 4A (no added drugs), illustrate that deleting the endogenous host CNB1 gene potentiates, or enhances, interactions between CNIc and calcineurin subunits CNA1, CNA2, CNA1 Δ C and CNA2 Δ C. Comparison of corresponding colonies in Figs. 4A and 4C shows the effects of FK506 on CNIc-CNA/CNA Δ C interactions. The drug enhances interactions under all except control conditions. The effect is most striking in yeast strains wild-type for the CNB1 subunit (e.g., compare 50a with 50b, and 51a with 51b).

The drug also markedly enhances, or potentiates CNIc-CNA/CNA Δ C interactions under conditions where the CNB1 subunit is overexpressed. Figure 5A shows the effect of overexpressing CNB1 on CNIc-CNA/CNA Δ C interactions in the absence of drug. Colonies expressing B1/YEp352 (53a, 55a-58a) have reduced signal as compared with controls expressing only YEp352 (52a, 54a). Inclusion of FK506 in the plating medium (Fig. 5C), however, enhances, or potentiates interactions in all colonies, except the negative controls (56).

The data presented in Figures 4A, 4C, 5A and 5C demonstrate that the interaction of CNIc with CNA and CNA Δ C is markedly enhanced by FK506. The interaction is also enhanced by deletion of CNB1 and diminished by overexpression of CNB1, and the inhibitory effect of CNB1 overproduction is overcome by the stimulatory effect of FK506.

Stated another way, inclusion of a small molecule immunosuppressant (FK506) potentiates an interaction between two fusion hybrid proteins, where one of the two proteins contains an (A) subunit of calcineurin, and the other protein contains a CNI polypeptide. The potentiation is particularly strong when the cell is further modified to cause overexpression of a "B" subunit of calcineurin by said cell (e.g., the expresses B1/YEp352).

In the present case, a yeast cell is modified to cause overexpression of a "B" subunit of calcineurin (CNB1) by transforming the cell with B1/YEp352 (construction described below). A cell may be modified to cause overexpression of a "B" subunit of calcineurin in other ways as well, such as, for example, transformation with other types of expression vectors encoding a "B" subunit of calcineurin, or treatment with a substance that upregulates a promoter controlling expression of an endogenous (B) subunit of calcineurin.

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In light of the effects of FK506 on CNIc-CNA/CNA Δ C interactions, two other immunosuppressants, cyclosporin A and rapamycin, were examined in similar experiments. The results of these experiments are shown in Figures 6A-6F and 7A-7H.

The data presented in Figures 6A and 6C are essentially equivalent to those presented in Figures 4A and 4C, respectively. Data shown in Figure 6E demonstrate that like FK506, the immunosuppressant cyclosporin A is also effective to enhance interaction of CNIc with CNA and CNAΔC. Both FK506 and cyclosporin A are known to exert their immunosuppressive effects through calcineurin (Cyert).

Similarly, data shown in Figures 7A and 7C support data in Figures 5A and 5C, and further, demonstrate that there is no detectable interaction between FK506 binding protein (FKBP) and CNIc. Results shown in Fig. 7E demonstrate that cyclosporin A has a similar effect to FK506 in cells overexpressing CNB1 — that is, it enhances the interactions between CNIc and CNA/CNAAC.

In contrast, data presented in Fig. 7G show that the immunosuppressant rapamycin has no detectable effect on CNIc-CNA/CNA Δ C interactions (compare Fig. 7G with Fig. 7A).

Taken together, the data presented in Figs. 6A, 6C, 6E 7A, 7C, 7E and 7G show that like FK506, cyclosporin A (CsA), but not rapamycin, enhances the interaction of CNIc with CNA and CNAAC, and that CNIc doesn't interact with FKBP with or without FK506.

Example 4 describes experiments to assess effects of CNIc on FKBP/FK506 binding to calcineurin. Figure 8A presents exemplary data from studies to assess the effect of CNI on FKBP-mediated FK506 interactions with CNA2. The legend for Fig. 8A is shown in Figure 8B. Data in Figure 8A demonstrate that, in the absence of FK506, FKBP and CNA2 show no detectable interaction (84). In the presence of FK506, however, the proteins interact (85), presumably because FK506 forms a complex with FKBP, which then binds CNA2 (Cyert).

Data in Figure 8A further show that, in the absence of FK506, CNI has no effect on the lack of interaction between FKBP and CNA2 (86). In the presence of FK506 (87), however, CNI potentiates, or enhances the binding between FKBP and CNA2 (compare the intensity at 87 with that at 85). This effect suggests that CNI and similar compounds may be employed to increase the sensitivity of calcineurin to immunosuppressant drugs that act on it, and in this way, decrease the amount of the immunosuppressant required for a particular level of immunosuppression.

Experiments illustrated in Figures 9A and 9B demonstrate that FK506 has little or no effect on the binding of CNA to CNB1. The legends for Fig. 9A and 9B are shown in Figures 9B and 9D, respectively. The data in Fig. 9A show that overexpression of the full-length CNI

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clone markedly enhances the FK506-dependent interaction of FKBP with CNA, although it doesn't affect the interaction between CNA and CNB1.

Example 5 presents co-immunoprecipitation of CNIc (carrying an HA epitope tag) and CNA. Immunoprecipitation is carried out with anti-HA monoclonal antibody and the immune complex, resolved by SDS-PAGE, is detected with anti-CNA2 polyclonal antibody and visualized with goat anti-rabbit antibody using the "ECL" method (Amersham, Arlington Heights, IL). The results, shown in Figure 10, demonstrate that CNIc is capable of binding to CNA2 tightly enough for the complex to be co-immunoprecipitated. Similar methods may be employed to isolate a CNI analog from other cell sources, including mammalian (specifically human).

Example 6 describes yeast RNA blots hybridized with a CNIc probe. Exemplary data are shown in Figure 11. A single message of approximately 2.9 kb is detected. The data indicate that CNI is an expressed gene encoding a 2.9 kb message in yeast.

Example 8 details the construction of cni null mutants. The null mutants are employed to assess if CNI is required for viability in yeast, and to test hygromycin B sensitivity. The experiments indicate that CNI is not essential for viability, since CNI deletion mutant strains can survive, but that CNI deletions render a host more resistant to hygromycin B. The effect is particularly pronounced in both MCY300-1 (cna1 cna2) and DD12 (cnb1), suggesting that CNI functions as a suppressor of CN mutant's sensitivity to hygromycin B.

It will be understood that all of the above methods and experimental manipulations are amenable to being done with interacting polypeptides from organisms other than yeast. In particular, calcineurin subunits, CNI polypeptides, immunophilins and the like may be of mammalian origin, e.g., human origin.

25 VII. UTILITY

Methods and compositions of the present invention may be applied in a number of different ways. Following the guidance presented herein, one of skill in the art may isolate nucleic acids encoding additional CNI polypeptides, for example, a human CNI polypeptide.

In one approach, a yeast strain carrying a mutation of the CNI gene, e.g., a deletion, is used to clone heterologous sequences (e.g., human sequences) by complementation. A library of genomic DNA or, preferably, cDNA from an organism (e.g., human) and tissue (e.g., lymphocyte cells) of choice is cloned into a vector that can be maintained in yeast. Preferably, the vector contains a yeast promoter effective to express the heterologous sequences in yeast cells. Several heterologous libraries suitable for expression in Saccharomyces

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cerevisiae containing DNA from S. pombe (Beach, et al.) and Drosophila have been constructed.

The library is transformed into a suitable yeast strain carrying a *cni* mutation, and transformants are selected using a suitable complementation assay. For example, transformants may be screened for increased hygromycin sensitivity, as experiments described herein indicate that *cni* deletion mutants possess a decreased sensitivity to hygromycin B (Example 8). The screen may be made more effective by using a yeast strain that is hypersensitive to hygromycin B, such as a strain deficient for a subunit of calcineurin (Example 8).

Alternatively, human CNI DNA sequences may be isolated by directly screening a library, e.g., a lymphocyte cDNA library, for clones hybridizing with a yeast CNI nucleic acid probe. The generation of an exemplary yeast CNI nucleic acid probe is described in Example 1.

In another approach, particularly advantageous for isolating sequences expressed at low levels, a CNI nucleic acid probe may be used to screen a genomic library, e.g., a human genomic library, to isolate a sequence that may be used to design probes or primers that may match the target sequence better that the yeast sequence. Such primers may be used with, for example, PCR, to isolate longer fragments from a tissue-specific library.

In yet another approach, an antibody generated against CNI polypeptide is used to immunoprecipitate a CNI polypeptide from an organism and/or tissue of choice. The protein may then be micro-sequenced, and the sequence utilized to design degenerate primers useful for isolating a cDNA.

CNI polypeptides of the present invention, particularly CNI fragments that retain a desired binding activity, may be used as lead compounds useful for the development of small molecules having cellular functions similar to those of the CNI-polypeptides, that is, molecules effective to enhance immunosuppressive effects of calcineurin-targeted immunosuppressants by potentiating an interaction of an immunophilin with calcineurin.

CNI-polypeptides of the present invention may also be employed in a method of increasing sensitivity of cells to calcineurin-affecting immunosuppressant drugs. In this method, a CNI-polypeptide is introduced into the cell typically prior to or at the same time as contacting the cell with an immunosuppressant drug, such as FK506. The polypeptide may be delivered by any suitable means effective to deliver polypeptides to selected cells,

Alternatively, nucleic acids encoding CNI polypeptides may be used in appropriate expression vectors as a genetic therapy tool to potentiate the immunosuppressive effects of

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calcineurin-targeting immunosuppressant drugs. The vectors may be targeted to selected cells, such as T-cells, to increase their sensitivity to a given systemic dose of an immunosuppressant.

Another utility of the present invention includes methods of screening for substances that up-regulate expression of CNI polypeptides, *i.e.*, substances that affect transcription. Such substances are useful for sensitizing cells to immunosuppressant drugs. In this method, the CNI promoter can be attached to a gene that functions as a selectable marker (for use in genetic selections to screen test substances) or to a reporter gene (for use in evaluating the effect on CNI transcription by test substances).

In another aspect of the present invention, the CNI-polypeptides, for example, mammalian homologue polypeptides of CNI, have potential use as therapeutic agents for both human and veterinary use. For example, CNI-polypeptides may be used in a method of enhancing immunosuppression in a test subject. In this method, the CNI-polypeptide is administered to the subject in a pharmaceutically-acceptable formulation and at a concentration effective to potentiate the interaction of an immunosuppressant/immunophilin complex with a subunit of calcineurin. The method may also include contacting the CNI-polypeptide with a cell under conditions effective to permit uptake of the protein into the cell in order to increase sensitivity of the cell to immunosuppressants. A CNI polypeptide used in such methods may be modified to be more suitable for administration or to be more effective in a cell. For example, a CNI polypeptide may be modified to eliminate PEST motifs, which are typically found in proteins with short half-lives, to extend the effective lifetime of the polypeptide in the target cell.

The following examples illustrate, but in no way are intended to limit the present invention.

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MATERIALS AND METHODS

Unless indicated otherwise, chemicals and reagents were obtained from Sigma Chemical Company (St. Louis, MO) or Mallinckrodt Specialty Chemicals (Chesterfield, MO), restriction endonucleases were obtained from New England Biolabs (Beverly, MA), and other modifying enzymes and biochemicals were obtained from Pharmacia Biotech (Piscataway, NJ) or Boehringer Mannheim (Indianapolis, IN). FK506 was obtained from Fujisawa USA, Inc. (Deerfield, IL), cyclosporin A was obtained from Sandoz (Basel, Switzerland), and rapamycin was obtained from Wyeth-Ayerst (Princeton, NJ). Materials for media for yeast growth and culture were obtained from DIFCO (Detroit, MI). Unless otherwise indicated, manipulations

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of yeast, bacteria, nucleic acids, proteins and antibodies were performed using standard methods and protocols (e.g., Guthrie, et al., Sambrook, et al., Ausubel, et al., Harlow, et al., and Rose, et al.).

A. Buffers

Z buffer: 60 mM Na₂HPO₄-7H₂O, 40 mM NaH₂PO₄-H₂O, 10 mM KCl, 1 mM MgSO₄-7H₂O and 50 mM β -mercaptoethanol (pH 7.0).

B. Plasmids, Libraries and Yeast Strains

Plasmids pGBT9 (GBT), carrying GAL4 DNA-binding domain (amino acid residues 1-147; G4BD) and TRP1, and pGAD (GAD), carrying GAL4 activation domain (amino acid residues 768-881; G4AD) and LEU2; three pGAD libraries carrying fusions between G4AD and yeast genomic Sau3AI partial-digest fragments in each frame; and the yeast GAL1-lacZ reporter strain SFY526 (Y526; MATa ura3-52 ade2-101 lys2-801 his3-200 trp1-901 leu2-3,112 can1 gal4-542 gal80-538 URA3::GAL1-lacZ) were obtained from Stanley Fields (State University of New York at Stony Brook, Stony Brook, NY; Chien, et al., Bartel, et al.). The libraries were constructed with linkers between the GAL4 activation domain and the Sau3AI fragments. The sequences of the linkers were 5'-ATCG-3'for the first library, 5'-ATCCG-3'for the second library, and 5'-ATCCCG-3'for the third library. In this way, the yeast genomic Sau3AI fragments were cloned in all three reading frames relative to G4AD.

Plasmids pAS2 (AS) carrying G4BD and TRP1, and pAS-lamin (AS-lamin) containing a sequence encoding a G4BD-lamin C fusion; and yeast reporter strains Y190 (MATa ura3-52 ade2-101 his3-Δ200 trp1-901 leu2-3,112 cyh2Δ, gal4Δ gal80Δ URA3::GAL-lacZ LYS2::GAL-HIS3), a derivative of Y153 carrying dual indicator genes (GAL-lacZ and GAL-HIS3), and Y187 (MATα ura3-52 ade2-101 lys2-801 his3-200 trp1-901 leu2-3,112 gal4Δ gal80Δ URA3::GAL-lacZ) carrying GAL-lacZ reporter were obtained from Stephen Elledge (Baylor College of Medicine, Houston, TX; Durfee, et al.). Yeast strain Y153b1 (cnb1::ADE2) was derived from Y153.

E coli strain JBe181 (leuB600 trpC9830) was obtained from Ira Herskowitz (University of California at San Francisco, San Francisco, CA). Protease-deficient yeast strain BJ2407 (Guthrie, et al.) was obtained from the Yeast Genetic Center (University of California at Berkeley, Berkeley, CA).

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C. GAL4-Calcineurin Fusions

GAL4-calcineurin (GAL4-CN) fusions, GBT-A1 (G4BD-CNA1), GBT-A2 (G4BD-CNA2), GBT-B1 (G4BD-CNB1), GAD-A1 (G4AD-CNA1), GAD-A2 (G4AD-CNA2), and GAD-B1 (G4AD-CNB1) were constructed as follows. Plasmids containing inserts encoding CN subunits CNA1 (SEQ ID NO:9; Cyert, et al., 1991), CNA2 (SEQ ID NO:11; Cyert, et al., 1991) and CNB1 (SEQ ID NO:13; Cyert, et al., 1992) were subjected to site-directed mutagenesis (Kunkle) to introduce a BamHI site just upstream of each subunit's initiation codon in the second reading frame. DNA prepared from the mutated plasmids was digested with BamHI and Xhol, and the resulting BamHI-Xhol fragments, each containing a full-length coding sequence, were cloned into GBT or GAD that had been cut with BamHI and Sal I. The resulting plasmids encoded in-frame fusions of the CN subunits with G4BD or G4AD.

Plasmids encoding CNA protein variants with truncated C-termini (GBT-A1 Δ C, GBT-A2 Δ C) were constructed by introducing stop codons after amino acid residues 509 (CNA1) and 502 (CNA2). The 44-residue deletion in GBT-A1 Δ C removed the autoinhibitory domain of CNA1, while the 102-residue deletion in GBT-A2 Δ C removed both the autoinhibitory and the calmodulin-binding domains of CNA2 (Cyert, et al., 1991).

Plasmid GBT-FKBP, containing an FK506 binding protein (FKBP) gene fused to the GAL4 binding domain, was constructed by introducing a Bg/II site upstream of the initiation codon and a BamHI site downstream of the stop codon of FKBP12 (Heitman, et al.) and ligating the Bg/II-BamHI fragment into GBT cut with BamHI.

Plasmid B1/YEp352 was constructed to contain the full coding sequence of CNB1 (SEQ ID NO:13, Cyert, et al., 1992) as a 1.4 kb BamHI-EcoRI fragment encompassing the sequence presented as SEQ ID NO:13 (812 bp; contains the coding sequence), in the multicopy plasmid YEp352(HIS), which is derived from YEp352 (URA) (Hill, et al.).

Plasmid CNI/YEp352(HIS) (also referred to as CNIH) was constructed by ligating a 3.16 kb MunI-EcoRV fragment, containing the full coding sequence of CNI, from plasmid CNI7.1 (construction described below) into YEp352(HIS) cut with EcoRI and Smal. Plasmids CNI/YEp352(TRP) (also referred to as CNIT) and CNI/YEp352(URA) (also referred to as CNIU) were similarly constructed using the 3.16 kb fragment and YEp352(TRP) or YEp352(URA), respectively (Hill, et al.).

Plasmids A1/YEp351 and A2/YEp352 were constructed to contain the full coding sequences of CNA1 (SEQ ID NO:9, Cyert, et al., 1991) in YEp351 (Hill, et al.) and CNA2 (SEQ ID NO:11, Cyert, et al., 1991) in YEp351 and YEp352, respectively. A1/YEp351 was constructed by ligating a 2.9 kb SacI-HindIII fragment from clone CNA1 (Cyert, et al., 1991)

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into YEp351(HIS) cut with SacI and HindIII. A2/YEp352 was similarly constructed by ligating a 3 kb SpeI-HindIII fragment from clone CNA2 (Cyert, et al., 1991) into YEp352(HIS) cut with XbaI and HindIII.

All GAL4-CN fusions were verified by DNA sequencing (Sanger, et al.) using "SEQUENASE 2.0" sequencing kits (United States Biochemical, Cleveland, OH), and were subjected to the following tests. The functionality of the fusion proteins was assayed by determining whether they could complement the appropriate cn mutant phenotypes, using assays to measure the sensitivity to pheromone and Mn²⁺ (Reneke, et al., Cyert, et al., 1991). All of the GAL4-CN fusions were functional in this assay.

The fusion proteins were also tested for their ability to activate the reporter gene in the absence of the complementary GAL-4 domain fusion (i.e., in the presence of the complementary GAL4 domain not fused to a second protein, for example, G4BD-A1 vs G4AD) using the two-hybrid interaction assay described below. Only GBT-B1 and GAD-A1 were able to activate the reporter gene at low levels without the complimentary GAL-4 domain fusion — assays with the other fusion proteins in the absence of the complimentary GAL-4 domain fusion showed no detectable levels of expression.

The two-hybrid interaction assay was also used to test the ability of the fusions to interact specifically with another fusion containing complimentary GAL4 and CN domains (e.g., G4BD-A1 interacting with G4AD-B1). All CN hybrids were able to react specifically and result in an activation of the reporter gene that was clearly detectable above background. The high specificity witnessed in these experiments indicates that the GAL4 two-hybrid system can reliably be used to assay interactions between CN and other proteins.

D. Yeast GAL4 Two-Hybrid System for Detecting Protein-Protein Interaction

In the library screen, described in more detail in Example 1A, the yeast strain Y190, harboring the hybrid plasmid carrying the GAL4 binding domain fused to the A1 subunit of calcineurin (G4BD-CNA1), was transformed with fusion libraries carrying yeast genomic DNA Sau3AI fragments fused to the GAL4 activation domain. Transformants that were able to express the reporter genes, *i.e.*, able to grow on -His + 3-AT and to score blue in β -gal assay, were selected as candidate positives. These candidate positives potentially contain library DNA fragments encoding proteins that physically interact with CNA1.

In another application described herein, the two-hybrid system was used to test for interactions between CNA (fused to one of the GAL4 domains) and CNB1 (fused to the other GAL4 domain), and between CNA and FKBP. Additional experiments tested a clone, CN1c.

isolated using the library screen, against a series of proteins fused to the complementary GAL4 domain under various conditions to test whether CN1c interacts with CNA subunits, and if so, how the interactions are affected by various conditions.

E. Color Development (β-gal) Assay

Yeast reporters harboring both G4BD and G4AD fusions (and a third non-fusion plasmid in some cases) were monitored for β -gal activity as follows. Purified yeast transformants were patched onto selective plates with or without other test reagents. After growing 3 days at 30°C, colonies were lifted onto nitrocellulose filters, permeabilized in liquid nitrogen as above, placed on Whatman No. 1 paper in petri dishes containing 0.1% X-Gal in Z buffer (see above), and incubated at 30°C for 12 hours. Blue color begins to appear in positive colonies between about one half and ten hours into the incubation period.

Exemplary images obtained using the color development assay are presented in Figures 2A, 3A, 4A, 4C, 5A, 5C, 6A, 6C, 6F, 7A, 7C, 7E, 7G, 8A, 9A and 9C.

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F. Growth Assay

A growth assay, applicable to yeast strains Y190 and Y153b1 which carry both GAL-HIS3 and GAL-lacZ reporters, was sometimes used as a complement to the color assay described above. Yeast transformants were streaked onto selective plates containing 40-50 mM 3-AT and no Histidine, and incubated at 30°C for 3-7 days. Growth (corresponding to the level of HIS3 expression) was monitored as an indicator of the interaction between fusion proteins. In cases where both assays were used, the amount of cell growth typically correlated well with the color intensity in the β -gal assay.

25 G. Yeast Growth, Drug Treatment

Yeast were typically grown in YPD (rich non-selective) or synthetic complete (SC) medium with selected component drop-outs, depending on the plasmid introduced, following standard procedures (Sherman, et al., Ausubel, et al.).

Experiments utilizing treatment with drugs or additives were performed by including the drug or additive in the medium. For plating, the agar was autoclaved, allowed to cool to 50°C, and the drug or additive was added before pouring the plates. Unless otherwise indicated, drugs and additives were added to result in the following final medium concentrations: FK506: 1 μ g/ml, cyclosporin A: 10 μ g/ml, rapamycin: 10 ng/ml, and hygromycin B: 40 μ g/ml.

H. Antibodies

Polyclonal and monoclonal antibodies, for use in the present invention, can be prepared by standard methods (Harlow, et al.) utilizing the CNI polypeptides of the present invention, for example, a substantially purified CNI/ β -galactosidase fusion protein (Example 9). Antibodies can also be generated by recombinant techniques (Cabilly, et al.; Better, et al.; Skerra, et al.). In addition to whole antibody molecules, antibody fragments retaining the immunological specificity of the whole antibody may also be used in the practice of the present invention (e.g., Fab and F(ab')₂ fragments of IgG (Pierce Chemical, Rockford, IL)). The antibodies can be purified by standard methods to provide antibody preparations which are substantially free of serum proteins that may affect reactivity (e.g., affinity purification (Harlow et al.)).

EXAMPLE 1 Isolation of CNIc

15 A. Library Screening

Yeast strain Y190 was transformed with pGBT-CNA1 TRP1 (GBT-A1) hybrid plasmid using the transformation protocol described by Schiestl, et al. Transformants were selected, colony purified, and a single transformant was selected to make (Y190 GBT-A1)-competent cells, following the procedure described in Guthrie, et al..

The three pGAD yeast fusion libraries described above, carrying fusions between G4AD and yeast genomic DNA Sau3Al fragments in each reading frame, were then used to transform (Schiestl, et al.) the Y190 GBT-A1-containing cells. Transformants were plated onto SC-Trp-Leu-His plates containing 40 mM 3-aminotriazole (3-AT; Sigma Chemical Co., St. Louis, MO) and incubated at 30°C for 6 days to screen for HIS⁺ colonies (Durfee, et al.).

His⁺ colonies were replica plated onto nitrocellulose filters (Schleicher & Schuell, Keene, NH), frozen in liquid nitrogen for approximately 30 seconds, and incubated at 30°C for 12 hours with Z buffer (see above) containing the chromogenic substrate X-Gal (0.1%) to assay β -gal activity (Breeden, et al.).

Candidate positive (blue) colonies were re-streaked for single colonies. Single colonies were purified and retested using the above protocol. Colonies which reproducibly tested positive were screened using PCR with primers directed against the internal portion of GAL-4 (i.e. the portion between the DNA binding domain and the activation domain). The sequences of the primers, G4-PCR-A and G4-PCR-B, are given as SEQ ID NO:22 and SEQ ID NO:23,

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respectively. Colonies yielding a PCR product were identified as containing intact GAL4, and were eliminated.

The GBT-A1 TRP⁺ plasmid was eliminated by growing in Trp⁺ liquid media for 2-3 days, plating on -Leu media and then replica-plating on -Leu and -Trp plates to identify and eliminate colonies that had lost the GBT-A1 plasmid, yet still gave a positive signal.

Plasmid DNA was extracted from the remaining Leu⁺ candidates. The plasmid DNA was transformed into *E coli* JBe181 and plated on -Leu media to select for library plasmids. The library plasmids isolated by this method were introduced back to the yeast reporter strains either alone or with test G4BD fusions: GBT, GBT-A1, and AS-lamin.

A parallel specificity assay was conducted by mating. Candidate strains, as described above, were 3-AT growth positive and X-gal positive when both the library and GBT-A1 plasmids were present. After elimination of the GBT-A1 plasmid from these strains, strains that were Leu* Trp 3-AT growth and β -gal were mated to the following strains: Y187 (MAT α) carrying GBT, GBT-A1, or AS-lamin, and the diploids were assayed.

Among the 3-AT positive, β -gal positive candidates identified by the secondary screening method just described, one clone (III-21S, later termed GAD-CNIc) was specifically positive in conjunction with GBT-A1 in both the transformation assay and the mating assay.

B. <u>Sequence of CNIc</u>

Clone III-21S was sequenced as above. The sequence is presented herein as SEQ ID NO:1, and a schematic representation of the clone is shown in Figure 1A. The Sau3AI library insert encodes 306 amino acids followed by a stop codon in frame with the coding sequence of GALA, but does not contain an in-frame ATG start site upstream of the stop codon. This is consistent with the fragment encoding a c-terminal portion of a complete gene product.

25 Accordingly, the clone was termed CNIc, with the lowercase "c" representing "c-terminal".

The stippled portion between GAL-4AD and CNIc in Figure 1A represents the linker discussed in Materials and Methods, above. Also indicated in the Figure are the approximate locations of in-frame start (ATG) and stop codons, and Sau3AI restriction sites.

30 C. Isolation of a Full Length Clone

A ³²P-labeled CNIc probe was generated from the 1.22 kb CNIc insert of clone III-21S by polymerase chain reaction (PCR) using primers represented as SEQ ID NO:7 and SEQ ID NO:8. The probe was used to map the gene to the right arm of chromosome 11 by hybridization screening (Sambrook, *et al.*) a panel of λ clones (American Type Culture

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Collection (ATCC), Rockville, MD) spanning the entire yeast genome. Two clones, 70500 and 70590, gave positive hybridization signals. A phage lysate of clone 70500 in λ MG3 was obtained from the ATCC, was amplified, purified, restriction-mapped and used as a DNA source for subsequent cloning experiments.

The phage DNA was digested with SacI, yielding a 7.1 kb fragment containing the entire CNI gene. This fragment was cloned into "BLUESCRIPT SK" (Stratagene, La Jolla, CA) cut with SacI, yielding plasmid CNI7.1. Plasmid CNI7.1 was digested with MunI and EcoRV, releasing a 3.16 kb fragment containing the entire coding sequence of CNI. The 3.16 kb fragment was then cloned into each of YEp352(HIS), YEp352(TRP), YEp352(URA), and "BLUESCRIPT SK", each cut with EcoRI and SmaI, yielding plasmids CNIH, CNIT, CNIU and CNI3.2, respectively. The sequence of the 3.16 kb MunI/EcoRV fragment is encompassed by the 3.5 kb sequence presented as SEQ ID NO:4. The MunI site is at nucleotide 100 of SEQ ID NO:4, and the EcoRV site is at nucleotide 3263 of SEQ ID NO:4.

A schematic diagram of the sequence presented as SEQ ID NO:4 is shown in Figure 1B. This sequence contains the 3.16 kb MunI/EcoRV fragment used in many of the experiments described herein (depicted in Figure 1B as the portion between the MunI and EcoRV sites), which contains the entire 2.75 kb coding sequence of CNI (SEQ ID NO:6; graphically indicated in Figure 1B by the locations of the "ATG" and "Stop" codons).

Figure 13 also shows the location of certain features of the sequence. For example, "PEST" motifs (Rogers, et al., Dice) are indicated by bars over the corresponding sequence.

A search of known DNA and protein sequences turned up no obvious matches or homologies to genes in other organisms. Accordingly, CNI may represent a new type of calcineurin-binding protein.

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EXAMPLE 2

Binding of CNIc to Calcineurin

Y190 yeast carrying the plasmids indicated below were assayed for β -gal activity by color development assay described above to determine the specificity of binding of CNIc to subunits of calcineurin.

Exemplary data, in the form of images of filters having yeast colony replicas that had undergone the β -gal color development assay are shown in Figure 2A. The legend for Fig. 2A is shown in Figure 2B. Numbers in the legend refer to locations of yeast colonies expressing particular combinations of plasmid constructs. The constructs are as follows: 20:GBT-A1 and

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GAD-B1, 21:GBT-A1 and GAD-CNIc, 22:GBT-A1 and GAD-CNIc, 23:GBT-A1 and GAD-CNIc 24:GBT-A2 and GAD-CNIc 25:AS-lamin and GAD-CNIc 26:GBT and GAD-CNIc.

Yeast colonies used in the assay were derived by several different methods. Those at location 22 were purified colonies from the original library screen, those at 21 were colonies transformed with mini-prep DNA of the isolated GAD-CNIc plasmid, and the remaining colonies (23, 24, 25 and 26) were transformed with maxi-prep (Qiagen, Chatsworth, CA) DNA of GAD-CNIc.

A comparison of the intensities of the blue β -gal reaction product indicates that CNIc interacted strongly with CNA1 regardless of the source of the CNIc plasmid DNA (20, 21, 22 and 23), and somewhat less strongly with CNA2 (24). Neither cells containing AS-lamin with GAD-CNIc (25), nor cells containing only GBT with GAD-CNIc (26) showed a detectable signal above background. Two subunits of calcineurin (CNA1, CNB1) known to interact were used as a positive control for the assay (20).

In summary, the data above show that CNIc interacted specifically with CNA1 and CNA2, but not with G4BD or lamin C.

A similar set of experiments was conducted using constitutively-active CNA subunits, as well as calcineurin subunit CNB1. As described in Materials and Methods, above, CNA1ΔC and CNA2ΔC were each missing a C-terminal portion of the protein containing an autoinhibitory domain. Exemplary results from these experiments are shown in Figure 3A. The legend for Fig. 3A is shown in Figure 3B. Locations of yeast colonies expressing specific constructs are as follows: 27:GBT and GAD-CNIc, 28:GBT-A1ΔC and GAD-CNIc, 29:GBT-A2ΔC and GAD-CNIc, 30:GBT-A1ΔC and GAD, 31:GBT-A2ΔC and GAD, 32:GBT-B1 and GAD-CNIc, 33:GBT-B1 and GAD, 34:GBT-A1ΔC and GAD-B1, and 35:GBT-A2ΔC and GAD-B1.

The data show that GBT-A1 Δ C and GAD-CNIc (28) gave a definite positive signal, while GBT-A2 Δ C and GAD-CNIc (29) was weaker, though still detectable above its background (i.e. GBT-A2 Δ C and GAD; 31). The signal from GBT-B1 and GAD-CNIc (32) was not detectable above vector background (GBT-B1 and GAD; 33). Positive controls GBT-A1 Δ C and GAD-B1 (34) and GBT-A2 Δ C and GAD-B1 (35) gave strong signals.

These data show that CNIc interacted specifically with CNA1ΔC and CNA2ΔC, but not with CNB1.

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EXAMPLE 3

Effects of Immunosuppressant Drugs on Binding of CNIc to Calcineurin in B1st, B1 Deletion and B1 Overproducing Yeast Strains

Three yeast strains carrying the plasmids indicated below were assayed for β-gal activity as above to determine if the immunosuppressant drugs FK506, cyclosporin A (CsA) and rapamycin affect the binding of CNIc to subunits of calcineurin. The experiments were performed in yeast strains wild-type for the CNB1 subunit, null for the CNB1 subunit, and in yeast transformed with a high efficiency expression vector containing DNA encoding the CNB1 subunit.

Exemplary data, in the form of filter images produced as above are shown in Figures 4A, 4C, 5A and 5C. Plates used to make the filters shown in Figs. 4A and 4C were replicas from one master plate, while plates used to make the filters shown in Figs. 5A and 5C were replicas from another plate. The plates used to generate filters shown in Figs. 4A and 5A were without FK506, while the plates used to generate filters shown in Figs. 4C and 5C contained $1 \mu g/ml$ FK506.

The legends for Figs. 4A and 4C are shown in Figures 4B and 4D, respectively. Since the imaged filters in Figs. 4A and 4C were from replica plates, corresponding locations on each filter contain material from the same yeast colonies. Accordingly, the locations are referred to by the same "base" numbers in the legends. To facilitate reference to a specific location on a specific filter, the base numbers are followed by a lowercase letter that is different for each of the individual filters. For example, in the present figure, "a" follows the base numbers to identify locations on the filter shown in Fig. 4A, while a "b" follows the base numbers to identify locations on the plate in Fig. 4C. This labeling scheme is used in other experiments detailed herein where multiple filter lifts are shown.

The interactions of various combinations of proteins expressed by constructs indicated below was studied in three yeast strains. Strain Y153b, at 36-40, is null for the CNB1 subunit of calcineurin. Strains Y190 (41-46) and Y526 (47-51) are wild-type for CNA1, CNA2 and CNB1.

Hybrid proteins expressed by colonies at specific locations are as follows: 36:GBT-A1 and GAD-CNIc, 37:GBT-A2 and GAD-CNIc, 38:GBT-A1ΔC and GAD-CNIc, 39:GBT and GAD-CNIc, 40:GBT-A2ΔC and GAD-CNIc, 41:GBT-A1 and GAD-CNIc, 42:GBT and GAD-CNIc, 43:GBT-A2 and GAD-CNIc, 44:GBT-A1ΔC and GAD-CNIc, 45:GBT-A1 and GAD-B1, 46:GBT-A2ΔC and GAD-CNIc, 47:GBT-A1 and GAD-CNIc, 48:GBT and GAD-CNIc,

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49:GBT-A2 and GAD-CNIc, 50:GBT-A1ΔC and GAD-CNIc, and 51:GBT-A2ΔC and GAD-CNIc.

Yeast strain Y526 was used for all experiments shown in Figs. 5A, 5B, 5C and 5D. The expression vector B1/YEp352(HIS) was not used in strains Y190 or Y153b1 because they are HIS⁺ in the absence of 3-AT.

The base numbers in Figures 5B and 5D correspond to locations of colonies expressing the following constructs: 52:GBT-A1, GAD-CNIc and YEp352, 53:GBT-A1, GAD-CNIc and B1/YEp352, 54:GBT-A2, GAD-CNIc and YEp352, 55:GBT-A2, GAD-CNIc and B1/YEp352, 56:GBT, GAD-CNIc and B1/YEp352, 57:GBT-A1ΔC, GAD-CNIc and B1/YEp352, and 58:GBT-A2ΔC, GAD-CNIc and B1/YEp352.

A comparison of data shown in Fig. 4A (no added drugs) shows the effect of deleting the endogenous host CNB1 gene on interactions between CNIc and calcineurin subunits CNA1, CNA2, CNA1ΔC and CNA2ΔC. Note that interactions in panels 36a-40a (CNB1 null strain) were all stronger (with the exception of the negative control in 39) than interactions in corresponding panels 41a-51a (strains wild-type for CNB1). This result indicates that interaction between CNIc and CNA subunits were enhanced by the deletion of the CNB1 subunit.

Comparison of corresponding panels in Figs. 4A and 4C shows the effects of FK506 on CNIc-CNA/CNA\(Delta\) Control interactions. The drug enhanced interactions under all except control (39, 42 and 48) conditions. The effect was most striking in yeast strains wild-type for the CNB1 subunit (e.g., compare 50a with 50b, and 51a with 51b).

The drug also markedly enhanced CNIc-CNA/CNA\(Delta\)C interactions under conditions where the CNB1 subunit was overexpressed. Figure 5A shows the effect of overexpressing CNB1 on CNIc-CNA/CNA\(Delta\)C interactions in the absence of drug. Colonies expressing B1/YEp352 (53a, 55a-58a) had reduced signal as compared with controls expressing only YEp352 (52a, 54a). Inclusion of FK506 in the plating medium (Fig. 5C), however, enhanced interactions in all colonies, except the negative controls (56).

Taken together, the above data demonstrate that the interaction of CNIc with CNA and CNA Δ C was markedly enhanced by FK506. The interaction was also enhanced by deletion of CNB1 and diminished by overexpression of CNB1, and the inhibitory effect of CNB1 overproduction was overcome by the stimulatory effect of FK506.

In light of the effects of FK506 on CNIc-CNA/CNADC interactions, two other immunosuppressants, cyclosporin A and rapamycin, were examined in similar experiments. The results of these experiments are shown in Figures 6A-6F and 7A-7H. Filters shown in

Figs. 6A-6F were from replica plates, as were those in Figs. 7A-7H. Colonies shown in Figs. 6A and 7A were plated without drugs; those in Figs. 6C and 7C were plated with FK506 (1 μg/ml), those in Figs. 6E and 7E with CsA (10 μg/ml), and those in Fig. 7G with rapamycin (10 ng/ml). Yeast strains used were as follows: In Figs. 6A-6F, panels 59-63 were Y153b1, 64-69 were Y190, and 70-74 were Y526. In Figs. 7A-7H, panels 77-83 were Y526, and panels 75 and 76 were Y190.

The base numbers in Figures 6B, 6D and 6F correspond to locations of colonies expressing the following constructs: 59:GBT-A1 and GAD-CNIc, 60:GBT-A2 and GAD-CNIc, 61:GBT-A1\Delta C and GAD-CNIc, 62:GBT and GAD-CNIc, 63:GBT-A2\Delta C and GAD-CNIc, 64:GBT-A1 and GAD-CNIc, 65:GBT and GAD-CNIc, 66:GBT-A2 and GAD-CNIc, 67:GBT-A1\Delta C and GAD-CNIc, 68:GBT-A1 and GAD-B1, 69:GBT-A2\Delta C and GAD-CNIc, 70:GBT-A1 and GAD-CNIc, 71:GBT and GAD-CNIc, 72:GBT-A2 and GAD-CNIc, 73:GBT-A1\Delta C and GAD-CNIc, and 74:GBT-A2\Delta C and GAD-CNIc.

The base numbers in Figures 7B, 7D, 7F and 7H correspond to locations of colonies expressing the following constructs: 75:GBT-FKBP and GAD, 76:GBT-FKBP and GAD-CNIc, 77:GBT-A1, GAD-CNIc and YEp352, 78:GBT-A1, GAD-CNIc and B1/YEp352, 79:GBT-A2, GAD-CNIc and YEp352, 80:GBT-A2, GAD-CNIc and B1/YEp352, 81:GBT, GAD-CNIc and B1/YEp352, 82:GBT-A1ΔC, GAD-CNIc and B1/YEp352, and 83:GBT-A2ΔC, GAD-CNIc and B1/YEp352.

The data presented in Figures 6A and 6C are essentially equivalent to those presented in Figures 4A and 4C, respectively. The constructs and yeast strains at corresponding locations were the same. As expected, the β -gal signal was also essentially equivalent between the two sets. Data shown in Figure 6E demonstrate that like FK506, the immunosuppressant cyclosporin A was also effective in enhancing interaction of CNIc with CNA and CNA Δ C. Both FK506 and cyclosporin A are known to exert their immunosuppressive effects through inhibition of calcineurin activity (Cyert).

Similarly, data shown in Figures 7A and 7C are essentially equivalent to those in Figures 5A and 5C, except that a top panel has been added in Figs. 7A-H. As above, the corresponding panels show the same constructs and yeast strains. The added panels (75 and 76) assessed the interaction of an FK506 binding protein (FKBP) with CNIc, and indicate that there were no detectable interactions between these proteins. Results in Fig. 7E demonstrate that cyclosporin A had a similar effect to FK506 in cells overexpressing CNB1 — that is, it enhanced the interactions between CNIc and CNA/CNAAC.

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In contrast, data presented in Fig. 7G show that the immunosuppressant rapamycin, which is known not to target calcineurin, had no detectable effect on CNIc-CNA/CNACC interactions (compare Fig. 7G with Fig. 7A).

Taken together, the above data show that like FK506, cyclosporin A (CsA), but not rapamycin, also enhanced the interaction of CNIc with CNA and CNAΔC. CNIc didn't interact with FKBP with or without FK506.

EXAMPLE 4

Effects of CNI on FKBP/FK506 binding to Calcineurin

Y526 cells, carrying the plasmids indicated below, were grown in -Trp-Leu-His liquid media with or without FK506 (1 μ g/ml) until OD₆₀₀ reached about 1.0. Approximately the same number of cells, calculated based on OD₆₀₀ and equivalent to 1 ml of an OD₆₀₀=1 suspension, was harvested from each culture, washed once with ddH₂O, centrifuged briefly, and the pellet was resuspended in 30 μ l ddH₂O and transferred onto a nitrocellulose filter. The filters were frozen in liquid nitrogen as described above, placed in a 8.5 cm petri dish containing a sheet of Whatman No. 1 paper (Whatman International LTD, Maidstone, UK) in 1.6 ml Z buffer containing 0.1% X-Gal, and incubated at 30°C for 8 hours.

Figure 8A presents exemplary data from studies to assess the effect of CNI overexpression on FK506-mediated FKBP interactions with CNA2. The legend for Fig. 8A is shown in Figure 8B. Locations of yeast colonies expressing specific constructs: 84:GBT-FKBP, GAD-A2 and YEp352, 85:GBT-FKBP, GAD-A2 and YEp352, 86:GBT-FKBP, GAD-A2 and CNI/YEp352, and 87:GBT-FKBP, GAD-A2 and CNI/YEp352. The cells at 85 and 87 were exposed to FK506, while those at 84 and 86 were not.

Data in Figure 8A demonstrate that, in the absence of FK506, FKBP and CNA2 showed no detectable interaction (84). In the presence of FK506, however, the proteins interacted (85), presumably because FK506 formed a complex with FKBP, which then bound CNA2 (Cyert).

Data in Figure 8A further show that, in the absence of FK506, CNI had no effect on the lack of interaction between FKBP and CNA2 (86). In the presence of FK506 (87), however, CNI potentiated, or enhanced the binding between FKBP and CNA2 (compare the intensity at 87 with that at 85). This effect suggests that CNI and similar compounds may be employed to increase the sensitivity of calcineurin to immunosuppressant drugs that act on it, and in this way, decrease the amount of the immunosuppressant required for a particular level of immunosuppression.

Experiments illustrated in Figures 9A and 9B demonstrate that CNI overproduction had little or no effect on the binding of CNA2 to CNB1, providing support for the specificity of the stimulatory effect that CNI overproduction had on the FK506-dependent binding of FKBP to calcineurin. The legends for Fig. 9A and 9B are shown in Figures 9B and 9D, respectively. Locations of yeast colonies expressing the following constructs: 88:GBT-A2, GAD-B1 and YEp352, 89:GBT-A2, GAD-B1 and YEp352, 90:GBT-A2, GAD-B1 and CNI/YEp352, and 91:GBT-A2, GAD-B1 and CNI/YEp352. The colonies at 89 and 91 were exposed to FK506, while colonies at 88 and 90 were not.

Taken together, the above data show that overexpression of the full-length CNI clone markedly enhanced the FK506-dependent interaction of FKBP with CNA, although it didn't affect the interaction between CNA and CNB1.

EXAMPLE 5

Co-Immunoprecipitation of CNIc and CNA

Yeast BJ2407 harboring AS-CNIc, which carries an influenza hemagglutinin (HA) epitope tag (Wilson, et al.), and GAD-A2 (lanes 1, 3) or A2/YEp352 (lanes 2, 5), and strain MCY300-1 (cnal cna2; lane 4) were grown in selective media to $OD_{600}=0.8$. The cells were harvested, lysed, and immunoprecipitated in the presence of 25 μ g/ml FK506 with anti-HA monoclonal antibody (obtained from M. Kirschner, Harvard University, Boston, MA; Wilson, et al.), following protocols described in Harlow, et al. The cell extracts (lanes 3-5) and the immune complex (lanes 1, 2) were resolved by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE; Laemmli) followed by western blot with a rabbit anti-CNA2 polyclonal antibody generated using standard methods (Harlow, et al.). Bound anti-CNA2 antibody was visualized with the "ECL" kit (Amersham, Arlington Heights, IL) using goat antirabbit antibody. Molecular weight markers are indicated on the right in kD.

The results demonstrate that CNIc was capable of binding to CNA2 tightly enough for the complex to be co-immunoprecipitated. This independent, biochemical assay confirmed the results described above obtained using the two hybrid protein interaction assay — that is, that CNIc physically interacted with and bound CNA subunits.

30 Cell extracts of BJ2407 harboring AS-CNIc, and GAD-A2 or A1/Yep351, and Y153b1 harboring AS-CNIc and GAD-A1 were subjected to SDS-PAGE followed by western blot with anti HA antibody. The results showed that CNIc was present at very low levels in vivo.

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The results are consistent with the observations that a limited amount of CNA2 was precipitated by anti-HA antibody recognizing the CNIc fusions, and that CNI contains PEST-like motifs, a feature of proteins with a short half-life in vivo (Rogers, et al.).

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EXAMPLE 6

Northern Blot of CNIc

Norther blots (e.g., Sambrook, et al.) of yeast total RNA were hybridized with a CNIc probe. Exemplary data are shown in Figure 11. 20 μ g yeast RNA from YPH499 (lane 1) and MCY300-1 (lane 2) was resolved in a formaldehyde-agarose gel, transferred onto "HYBOND N+" membrane (Amersham, Arlington Heights, IL), and hybridized with 5×10^6 cpm/ml probe of the CNIc insert (1.22kb). A single message of approximately 2.9 kb was detected in both strains at about the same level following an 18-hour exposure on XAR5 film (Eastman Kodak, Rochester, NY).

The data indicate that CNI was a physiologically expressed gene encoding a 2.9 kb message in yeast.

EXAMPLE 7

Chromosome Mapping of CNIc

A yeast chromosome blot obtained from the ATCC was hybridized with probe of the CNIc insert following the Southern hybridization procedure described in Sambrook, et al. A positive hybridization signal was obtained with two ATCC yeast genomic λ clones derived from chromosome 11. Clone 70500 had a relatively strong signal, while clone 70590 had a somewhat weaker one. A phage lysates of clone 70500 was ordered from the ATCC, amplified, purified, restriction-mapped, and used as a DNA source for cloning full length CNI (Example 1).

EXAMPLE 8

CNI null Mutants

1. Construction of cni Null Mutation

A 5', 1.8 kb Bg/II-HindIII and a 3', 0.9 kb XbaI-Bg/II fragment of CNI were ligated into pRS305(LEU2) (Sikorski, et al.). The resultant plasmid had a deletion of a 2 kb HindIII-XbaI fragment from the coding sequence of CNI. This cni::LEU2 mutant was introduced into the genomes of yeast haploid strains YPH499 (Sikorski, et al.), MCY300-1 (cnal cna2) and DD12 (cnbl-) (Cyert, et al., 1991, Cyert, et al., 1992) as well as two diploid strains.

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Leucine prototrophs were isolated at high frequency from all strains, and hybridization analysis confirmed that the *cni::LEU2* allele had replaced the CNI gene. The experiments indicate that CNI is not essential for viability, since CNI deletion mutant strains (even cni double mutants) can survive.

CM was deleted from three yeast strains: YPH499 (WT), MCY300-1 (cna1 cna2), and DD12 (cnb1), resulting in cni strains LHy499, LHy300 and LHy12, respectively. Cells representing four colonies of each *cni* knockout strain and two colonies of each parent strain were grown in liquid YPD (Sherman, *et al.*) to saturation. Same numbers of cells from each culture were then plated onto YPD+Hygromycin B (40 μ g/ml) and growth was monitored at 30°C.

CNI deletions in each strain rendered that strain more resistant to hygromycin B. The effect was particularly pronounced in both MCY300-1 and DD12, suggesting that CNI functions as a suppressor of CN mutant's sensitivity to hygromycin B. The data indicate that deletion of CNI results in higher resistance to hygromycin B.

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EXAMPLE 9

Isolation of CNI/B-Galactosidase Fusion Protein

A CNI coding sequence is cloned into the λ gt11 vector (Stratagene, La Jolla, CA). The coding frame is cloned in-frame to the β -galactosidase coding sequences present in λ gt11. Bacterial lysogens infected either with lambda phage gt11 or with gt11/CNI are incubated in 32°C until the culture reaches to an O.D. of 0.4. Then the culture is incubated in a 43°C water bath for 15 minutes to induce gt11 peptide synthesis, and further incubated at 37°C for 1 hour. Bacterial cells are pelleted and lysed in lysis buffer (10 mM Tris, pH 7.4, 2 % "TRITON X-100" and 1% aprotinin). Bacterial lysates are clarified by centrifugation (10K, for 10 minutes, Sorvall JA20 rotor) and the clarified lysates are incubated with Sepharose 4B beads conjugated with anti- β -galactosidase (Promega).

Binding and elution of β -galactosidase fusion proteins are performed according to the manufacturer's instruction. Typically binding of the proteins and washing of the column are done with lysis buffer. Bound proteins are eluted with 0.1 M carbonate/bicarbonate buffer, pH 10.

While the invention has been described with reference to specific methods and embodiments, it will be appreciated that various modifications and changes may be made without departing from the invention.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: The Board of Trustees of the Leland Stanford Junior University
- (ii) TITLE OF INVENTION: Calcineurin Interacting Protein Compositions and Methods
- (iii) NUMBER OF SEQUENCES: 23
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Dehlinger & Associates
 - (B) STREET: 350 Cambridge Avenue, Suite 250
 - (C) CITY: Palo Alto
 - (D) STATE: CA
 - (E) COUNTRY: USA
 - (F) ZIP: 94306
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk

 - (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE: 23-OCT-1995
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/328,322
 - (B) FILING DATE: 24-OCT-1994
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Sholtz, Charles K.
 - (B) REGISTRATION NUMBER: P38,615
 - (C) REFERENCE/DOCKET NUMBER: 8600-0151.41
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (415) 324-0880
 - (B) TELEFAX: (415) 324-0960
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1222 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: Sau3AI fragment containing CNIc coding sequence
 - (ix) FEATURE:

 - (A) NAME/KEY: CDS (B) LOCATION: 1..918

(xi)	SEQUENCE	DESCRIPTION:	SEQ	ID	NO: 1	:
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									AAA Lys 10							48
TAT Tyr	AAG Lys	CCC Pro	AAT Asn 20	CAG Gln	ATT Ile	CCA Pro	AGA Arg	GAA Glu 25	TTA Leu	ACT Thr	TCT Ser	CCT Pro	CAG Gln 30	GCG Ala	TTA Leu	96
CCA Pro	TTA Leu	TCG Ser 35	CCC Pro	ATC Ile	ACC Thr	TCA Ser	CCA Pro 40	ATT	CTC Leu	AAT Asn	TAC Tyr	CAA Gln 45	CCA Pro	TTA Leu	TCA Ser	144
AAC Asn	TCC Ser 50	CCG Pro	CCT Pro	CCA Pro	GAT Asp	TTT Phe 55	GAT Asp	TTT Phe	GAT Asp	CTA Leu	GCT Ala 60	AAG Lys	CGC Arg	GGC Gly	GCA Ala	192
									CCT Pro							240
									TAC Tyr 90							288
									TCT Ser							336
									TCT Ser							384
GAC Asp	GAA Glu 130	GAT Asp	GAC Asp	AAT Asn	GAT Asp	GGC Gly 135	GAT Asp	ATA Ile	GCA Ala	TCT Ser	GGT Gly 140	TTC Phe	AAC Asn	TTC Phe	AAG Lys	432
CTG Leu 145	TCA Ser	ACC Thr	AGT Ser	GCT Ala	CCG Pro 150	AGT Ser	GAG Glu	AAC Asn	GTT Val	AAT Asn 155	TCA Ser	CAC His	ACT Thr	CCT Pro	ATT Ile 160	480
TTG Leu	CAG Gln	TCT Ser	TTA Leu	AAC Asn 165	ATG Met	AGT Ser	CTT Leu	GAT Asp	GGG Gly 170	AGA Arg	AAA Lys	AAA Lys	AAT Asn	CGT Arg 175	GCC Ala	528
AGT Ser	CTA Leu	CAC His	GCA Ala 180	ACA Thr	TCA Ser	GTG Val	TTA Leu	CCT Pro 185	AGT Ser	ACA Thr	XTX Ile	AGA Arg	CAG Gln 190	AAC Asn	AAT Asn	576
CAG Gln	CAT His	TTC Phe 195	AAT Asn	GAC Asp	ATA Ile	AAC Asn	CAG Gln 200	ATG Met	CTA Leu	GGC	AGT Ser	AGT Ser 205	GAC Asp	GAA Glu	GAT Asp	624
GCC Ala	TTT Phe 210	CCC Pro	AAA Lys	AGC Ser	CAA Gln	TCA Ser 215	TTA Leu	AAT Asn	TTC Phe	AAT Asn	AAG Lys 220	AAA Lys	CTA Leu	CCA Pro	ATA Ile	672
CTT Leu 225	AAA Lys	ATT Ile	AAT Asn	GAT Asp	AAC Asn 230	GTC Val	ATA Ile	CAA Gln	TCA Ser	AAC Asn 235	AGC Ser	AAT Aen	AGT Ser	AAT Asn	AAC Asn 240	720
AGA Arg	GTT Val	GAT Aup	AAT Asn	CCA Pro 245	GAA Glu	GAT Asp	ACA Thr	GTG Val	GAT Asp 250	TCT Ser	TCA Ser	GTC Val	GAT Asp	ATT Ile 255	ACA Thr	768

														TGG Trp		816
														GTT Val		864
														GAA Glu		912
AGA Arg 305		TAAC	STACA	ATT /	TTT	CATT	C TO	CCGAC	CAGAJ	A TTC	CTAC	CAT	TTT	ACTT:	rg t	968
GTCC	TGT	AT T	CAAT	RAGTO	T AC	CAATA	TATI	r GGJ	CATI	KTTA	TAG	CATA	CAR	ATAT	ACACO	A 1028
TCAA	TCT	ATA (CATCO	CATA	C AC	TTG1	CGTA	AAC	CATA	CCC	TTT	CAAT	CAG	TACA	CGAT	TT 1088
AAAA	LAAAI	CAA C	CATGA	\TTAJ	C G1	TCAC	TTAC	CAJ	TGAC	CTT	ATT	(TTA	AGG (CTTG	CTTT	AG 1148
ATTI	TTC	CAA C	TCAP	ATTT1	T GI	TTT	TCTA	A ACC	CTTC	CAA	CCT	CATC	CA i	ACCT:	CTTC	C 1208
TTTG	CAAG	CA C	SATC													1222

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 306 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Asp Gln Ser Ser Asn Val Phe Ala Ser Lys Gln Leu Val Ala Asn Ile 1 5 10 15

Tyr Lys Pro Asn Gln Ile Pro Arg Glu Leu Thr Ser Pro Gln Ala Leu 20 25 30

Pro Leu Ser Pro Ile Thr Ser Pro Ile Leu Asn Tyr Gln Pro Leu Ser 40 45

Asn Ser Pro Pro Pro Asp Phe Asp Phe Asp Leu Ala Lys Arg Gly Ala 50 55 60

Ala Asp Ser His Ala Ile Pro Val Asp Pro Pro Ser Tyr Phe Asp Val 65 70 75 80

Leu Lys Ala Asp Gly Ile Glu Leu Pro Tyr Tyr Asp Thr Ser Ser Ser 85 90 95

Lys Ile Pro Glu Leu Lys Leu Asn Lys Ser Arg Glu Thr Leu Ala Ser 100 105 110

Ile Glu Glu Asp Ser Phe Asn Gly Trp Ser Gln Ile Asp Asp Leu Ser 115 120 125

Asp Glu Asp Asp Asn Asp Gly Asp Ile Ala Ser Gly Phe Asn Phe Lys 130 135 140

Leu Ser Thr Ser Ala Pro Ser Glu Asn Val Asn Ser His Thr Pro Ile 145 150 155 160

Leu Gln Ser Leu Asn Met Ser Leu Asp Gly Arg Lys Lys Asn Arg Ala Ser Leu His Ala Thr Ser Val Leu Pro Ser Thr Ile Arg Gln Asn Asn 180 Gin His Phe Asn Asp Ile Asn Gin Met Leu Gly Ser Ser Asp Glu Asp Ala Phe Pro Lys Ser Gln Ser Leu Asn Phe Asn Lys Lys Leu Pro Ile Leu Lys Ile Asn Asp Asn Val Ile Gln Ser Asn Ser Asn Ser Asn Asn 230 Arg Val Asp Asn Pro Glu Asp Thr Val Asp Ser Ser Val Asp Ile Thr Ala Phe Tyr Asp Pro Arg Met Ser Ser Asp Ser Lys Phe Asp Trp Glu 260 Val Ser Lys Asn His Val Asp Pro Ala Ala Tyr Ser Val Asn Val Ala 280 Ser Glu Asn Arg Val Leu Asp Asp Phe Lys Lys Ala Phe Arg Glu Lys 290 295 300

Arg Lys 305

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 918 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: CINc coding sequence
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GATCAAAGTA GCAATGTCTT CGCATCCAAA CAGCTGGTCG CAAACATTTA TAAGCCCAAT 60 CA ATTCCAA GAGAATTAAC TTCTCCTCAG GCGTTACCAT TATCGCCCAT CACCTCACCA 120 ATTCTCAATT ACCAACCATT ATCAAACTCC CCGCCTCCAG ATTTTGATTT TGATCTAGCT 180 AAGCGCGGCG CAGCCGATTC TCATGCTATT CCTGTGGATC CTCCATCATA TTTTGATGTA 240 TTANAGGCCG ATGGGATTGA ATTGCCATAC TACGATACAA GTTCATCTAA AATTCCTGAA 300 CTAAAACTAA ACAAATCTAG AGA ACATTG GCCAGCATTG AGGAGGACTC ATTCAATGGT 360 TGGTCTCAAA TTGATGACTT ATCCGACGAA GATGACAATG ATGGCGATAT AGCATCTGGT 420 TTCAACTTCA AGCTGTCAAC CAGT CTCCG AGTGAGAACG TTAATTCACA CACTCCTATT 480 TTGCAGTCTT TAAACATGAG TCTTGATGGG AGAAAAAAA ATCGTGCCAG TCTACACGCA 540



ACATCAGTGT	TACCTAGTAC	AATAAGACAG	AACAATCAGC	ATTTCAATGA	CATAAACCAG	600
ATGCTAGGCA	GTAGTGACGA	AGATGCCTTT	CCCAAAA CC	AATCATTAAA	TTTCAATAA	660
AAACTACCAA	TACTTAAAAT	TANTGATAAC	GTCATACAAT	CAAACAGCAA	TAGTAATAAC	720
AGAGTTGATA	ATCCAGAAGA	TACAGTGGAT	TCTTCAGTCG	ATATTACAGC	ATTTTATGAT	780
CCAAGAATGT	CATCAGATTC	CAAATTTGAT	TGGGAGGTAA	GCAAGAACCA	TGTTGACCCA	840
GCAGCCTACT	CGGTTAACGT	TGCTAGTGAA	AACCGTGTAC	TGGACGACTT	TAAGAAAGCA	900
TTTCGCGAAA	AGAGAAAA					918

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3500 base pairs

 - (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full CNI coding sequence
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 376..3120

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

CCGAACACTT CCTTCGAGAG AGTGCATTTT ACTATGTGAA CCAATTTTTC CTCTTTTTCC	60
GTTTGCAAGT TCACCTGAAA AACTGCTTAA CACTACTAGC AATTGCCCTA TTGTCGTACG	120
AGGACTTTGC CAAATGTATT CCCGGCTGTT TGTAGTATAT ATACGCAGAT ATATAATAGC	180
GCCGTCTTTT TACCTCTTTG AGCGAATTGC CAAATATTGA CTCTTTTGTC TTATTTCGCT	240
ATCCCCATCT TATCAAAAAT GGGAACAACT CGTTGAAATA AGAGACAAGC AACAAGAAAA	300
ACAACCAACA GAAAGTTCCA TTCCGCACAA ATACGCTGGA ATCCCATAGA ATATTGCTTC	360
TTCCTCTATG ACTAC ATG CTC CAA TTC AAT ACA GAA AAT GAT ACT GTA GCT Met Leu Gln Phe Asn Thr Glu Asn Asp Thr Val Ala	411
CCA GTG TTT CCC ATG GAG CAA GAT ATA AAT GCA GCA CCT GAT GCC GTC Pro Val Phe Pro Het Glu Gln Asp Ile Asn Ala Ala Pro Asp Ala Val 15 20 25	459
CCA CTG GTG CAG ACA ACA CTA CAA GTC TTT GTA AAG CTT GCC GAA Pro Leu Val Gln Thr Thr Leu Gln Val Phe Val Lys Leu Ala Glu	507
30 35 40	555



			Arg							CCG Pro	603
						Phe				ACA Thr	651
					Lys			TTT Phe 105		GTT Val	699
	Val							Ala		GCC GCC	747
								AAT Asn			795
								AAA Lys			843
								GAT Asp			891
								TCC Ser 185			939
								GAC Asp			987
								CGA Arg			1035
								CAT His			1083
								GCT Ala			1131
								GAT Asp 265			1179
								TTT Phe			1227
								TTT Phe			1275
								GCA Ala			1323
								CTT Leu			1371



					ACT												1419
					GAC Asp												1467
					ATC Ile 370												1515
					AGA Arg												1563
					AAT Asn												1611
					GCT Ala												1659
					CCT Pro												1707
					GGC Gly 450												1755
					AAG Lys												1803
					ATC Ile												1851
					TAC Tyr												1899
					CAT His												1947
					TAC Tyr 530												1995
					GCA Ala											,	2043
GAT Asp	TCC Ser	AAT Asn	ATT 11e 560	TTC Phe	TTC Phe	CCA Pro	AAA Lys	GAA Glu 565	GTT Val	CTT Leu	TCG Ser	TCT Ser	CCC Pro 570	GTT Val	CTT Leu	;	2091
TCA Ser	CCT Pro	AAC Asn 575	GTG Val	CAG Gln	AAG Lys	ATG Met	AAC Asn 580	ATT Ile	AGA Arg	ATA Ile	CCG Pro	TCT Ser 585	GAT Asp	CTT Leu	CCA Pro		2139
GTA Val	GTG Val 590	CGT Arg	AAT Asn	AGA Arg	GCT Ala	GAA lu 595	AGC Ser	GTA Val	AAG Lys	AAA Lys	AGC Ser 600	AAG Lys	TCA Ser	GAT Asp	AAT Asn		2187

ACC Thr 605	TCC Ser	AAG Lys	AAG Lys	AAT Asn	GAT Asp 610	CAA Gln	AGT Ser	AGC Ser	AAT Asn	GTC Val 615	TTC Phe	GCA Ala	TCC Ser	AAA Lys	CAG Gln 620	2235
CTG Leu	GTC Val	GCA Ala	AAC Asn	ATT Ile 625	TAT Tyr	AAG Lys	CCC Pro	AAT Asn	CAG Gln 630	ATT	CCA Pro	AGA Arg	GAA Glu	TTA Leu 635	ACT Thr	2283
						TTA Leu										2331
						TCC Ser										2379
						GAT Asp 675										2427
						AAG Lys										2475
						ATT Ile	_									2523
						GAG Glu										2571
						GAA Glu										2619
						TCA Ser 755										2667
						CAG Gln										2715
						CTA Leu										2763
ATA Ile	AGA Arg	CAG Gln	AAC Asn 800	AAT Asn	CAG Gln	CAT His	TTC Phe	AAT Asn 805	GAC Asp	ATA Ile	AAC Asn	CAG Gln	ATG Met 810	CTA Leu	GLY	2811
						TTT Phe										2859
AAG Lys	AAA Lys 830	CTA Leu	CCA Pro	ATA Ile	CTT Leu	AAA Lys 835	ATT Ile	AAT Asn	GAT Asp	AAC Aen	GTC Val 840	ATA Ile	CAA Gln	TCA Ser	AAC Asn	2907
AGC Ser 845	AAT Asn	AGT Ser	AAT Asn	AAC Asn	AGA Arg 850	GTT Val	GAT Asp	AAT Asn	CCA Pro	GAA Glu 855	GAT Asp	ACA Thr	GTG Val	GAT Asp	TCT Ser 860	2955

TCA Ser	GTC Val	GAT Asp	ATT Ile	ACA Thr 865	GCA Ala	TTT Phe	TAT Tyr	GAT Asp	CCA Pro 870	AGA Arg	AT Met	TCA Ser	TCA Ser	AT Asp 875	TCC Ser	3003
AAA Lys	TTT Phe	GAT Asp	TGG Trp 880	GAG Glu	GTA Val	AGC Ser	AAG Lys	AAC Asn 885	CAT His	GTT Val	GAC Asp	CCA Pro	GCA Ala 890	GCC Ala	TAC Tyr	3051
TCG Ser	GTT Val	AAC Asn 895	GTT Val	GCT Ala	AGT Ser	GAA Glu	AAC Asn 900	CGT Arg	GTA Val	CTG Leu	GAC Asp	GAC Asp 905	TTT Phe	AAG Lys	AAA Lys	3099
			GAA Glu				TAAC	STAC	ATT I	\TTT	rca t 1	C TO	CCGA	CAGAI	•	3150
TTG	TAC	CAT !	TTTAC	CTTT	GT GT	CCT	GTGA:	TC	NATA	STGT	ACAI	TAT	ATT (GGAC	ATTTTA	3210
TAGI	ATA?	CAA 2	ATATA	ACAC	CA TO	CAAT	CTAT	A CA	rcca:	TATC	ACT:	CTC	STA .	AAGA!	TATCCC	3270
TTTT	TAAT	rag :	TACA	GCGA:	TT A	AAAA	AATA	A CA	rgat'	TAAC	GTT	CAGT	rac .	CAAT	GAGCTT	3330
ATTI	TTAT	AGG (CTTG	CTTT	AG A	rttt:	CCA	A GT	CAAT	TTT	GTT:	TTTT	CTA .	ACGC:	TTGCAA	3390
CCT	CATC:	rca i	ACCT:	CTT	CC T	rtgc	NAGC	A GA	CTT	CGAA	ACC	ATCT	CGT	TTAT:	TCTCTC	3450
AATO	CTG	TTC (CCAC!	TTTC	AT C	ATCG'	CTG	G GA	AAAG!	TACC	GGT	AAGG	GCG			3500

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 915 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Leu Gln Phe Asn Thr Glu Asn Asp Thr Val Ala Pro Val Phe Pro

Met Glu Gln Asp Ile Asn Ala Ala Pro Asp Ala Val Pro Leu Val Gln

Thr Thr Leu Gln Val Phe Val Lys Leu Ala Glu Pro Ile Val Phe

Leu Lys Gly Phe Glu Thr Asn Gly Leu Ser Glu Ile Ala Pro Ser Ile

Leu Arg Gly Ser Leu Ile Val Arg Val Leu Lys Pro Asn Lys Leu Lys 65 70 75 80

Ser Ile Ser Ile Thr Phe Lys Gly Ile Ser Arg Thr Glu Trp Pro Glu

Gly Ile Pro Pro Lys Arg Glu Glu Phe Ser Asp Val Glu Thr Val Val 105

Asn His Thr Trp Pro Phe Tyr Gln Ala Asp Asp Gly Met Asn Ser Phe

Thr Leu Glu His His Ser Ser Asn Asn Ser Ser Asn Arg Pro Ser Met 140 130

Ser 145	Asp	Glu	Asp	Tyr	Leu 150	Leu	Glu	Lys	Ser	Gly 155	Ala	Ser	Val	Tyr	11e 160
Pro	Pro	Thr	Ala	Glu 165	Pro	Pro	Lys	Asp	Asn 170	Ser	Asn	Leu	Ser	Leu 175	Asp
Ala	Tyr	Glu	Arg 180	Asn	Ser	Leu	Ser	Ser 185	yab	Asn	Leu	Ser	Asn 190	Lys	Pro
Val	Ser	Ser 195	Asp	Val	Ser	His	Asp 200	Asp	Ser	Lys	Leu	Leu 205	Ala	Ile	Gln
Lys	Thr 210	Pro	Leu	Pro	Ser	Ser 215	Ser	λrg	Arg	Gly	Ser 220	Val	Pro	Ala	Asn
Phe 225	His	Gly	Asn	Ser	Leu 230	Ser	Pro	His	Thr	Phe 235	Ile	Ser	yab	Leu	Phe 240
Thr	Lys	Thr	Phe	Ser 245	Asn	Ser	Gly	Ala	Thr 250	Pro	Ser	Pro	Glu	Gln 255	Glu
Хsр	Asn	Tyr	Leu 260	Thr	Pro	Ser	Lys	Asp 265	Ser	Lys	Glu	Val	Phe 270	Ile	Phe
Arg	Pro	Gly 275	Asp	Tyr	Ile	Tyr	Thr 280	Phe	Glu	Gln	Pro	11e 285	Ser	Gln	Ser
Tyr	Pro 290	Glu	Ser	Ile	Lys	Ala 295	Asn	Phe	Gly	Ser	Val 300	Glu	Tyr	Lys	Leu
Ser 305	Ile	Хsр	Ile	Glu	Arg 310	Phe	Gly	Ala	Phe	Lys 315	Ser	Thr	Ile	His	Thr 320
Gln	Leu	Pro	Ile	Lys 325	Val	Val	Arg	Leu	Pro 330	Ser	Asp	Gly	Ser	Val 335	Glu
Glu	Thr	Glu	Ala 340	Ile	Ala	Ile	Ser	Lys 345	Asp	Trp	Lys	Asp	Leu 350	Leu	His
Tyr	Asp	Val 355	Val	Ile	Phe	Ser	Lys 360	Glu	Ile	Val	Leu	Asn 365	Ala	Phe	Leu
Pro	11e 370	Хsр	Phe	His	Phe	Ala 375	Pro	Leu	Asp	Lys	Val 380	Thr	Leu	His	Arg
11e 385	Arg	Ile	Tyr	Leu	Thr 390	Glu	Ser	Met	Glu	Tyr 395	Thr	Cys	Asn	Ser	As n 400
Gly	Asn	His	Glu	Lys 405	Ala	λrg	Arg	Leu	Glu 410	Pro	Thr	Lys	Lys	Phe 415	Leu
Leu	Ala	Glu	His 420	Asn	Gly	Pro	Lys	Leu 425	Pro	His	Ile	Pro	Ala 430	Gly	ser
λsn	Pro	Leu 435	Lys	Ala	Lys	Asn	Arg 440	Gly	Asn	Ile	Leu	Leu 445	Asp	Glu	Lys
Ser	Gly 450	Asp	Leu	Val	Asn	Lys 455	Asp	Phe	Gln	Phe	Glu 460	Val	Phe	Val	Pro
Ser 465	Lys	Phe	Thr	Asn	Ser 470	Ile	Arg	Leu	His	Pro 475	Asp	Thr	Asn	Tyr	Asp 480
Lys	Ile	Lys	Ala	His 485	His	Trp	Ile	Lys	Ile 490	Cys	Leu	Arg	Leu	Ser 495	Lys

Lys	Tyr	Gly	Asp 500	Asn	Arg	Lys	His	Ph e 505	Glu	Ile	Ser	Ile	As p 510	Ser	Pro
Ile	His	Ile 515	Leu	Asn	Gln	Leu	Cys 520	Ser	His	Ala	Asn	Thr 525	Leu	Leu	Pro
Ser	Tyr 530	Glu	Ser	His	Phe	Gln 535	Tyr	Cys	Asp	Glu	Asp 540	Gly	Asn	Phe	Ala
Pro 545	Ala	Ala	Asp	Gln	Gln 550	Asn	Tyr	Ala	Ser	His 555	His	Хsр	Ser	As n	11 e 560
Phe	Phe	Pro	Lys	Glu 565	Val	Leu	Ser	Ser	Pro 570	Val	Leu	Ser	Pro	As n 575	Val
Gln	Lys	Met	Asn 580	Ile	λrg	Ile	Pro	Ser 585	Asp	Leu	Pro	Val	Val 590	Arg	Asn
Arg	Ala	Glu 595	Ser	Val	Lys	Lys	Ser 600	Lys	Ser	Asp	Asn	Thr 605	Ser	Lys	Lys
Asn	Asp 610	Gln	Ser	Ser	Asn	Val 615	Phe	Ala	Ser	Lys	Gln 620	Leu	Val	Ala	Asn
11e 625	Tyr	Lys	Pro	Asn	Gln 630	Ile	Pro	Arg	Glu	Leu 635	Thr	Ser	Pro	Gln	Ala 640
Leu	Pro	Leu	Ser	Pro 645	Ile	Thr	Ser	Pro	11e 650	Leu	Asn	Tyr	Gln	Pro 655	Leu
Ser	Asn	Ser	Pro 660	Pro	Pro	Asp	Phe	Asp 665	Phe	Asp	Leu	Ala	Lys 670	Arg	Gly
Ala	Ala	λε ρ 675	Ser	His	Ala	Ile	Pro 680	Val	Asp	Pro	Pro	Ser 685	Tyr	Phe	Asp
Val	Leu 690	Lys	Ala	Asp	Gly	11e 695	Glu	Leu	Pro	Tyr	Tyr 700	Asp	Thr	Ser	Ser
Ser 705	Lys	Ile	Pro	Glu	Leu 710	Lys	Leu	λsn	Lys	Ser 715	Arg	Glu	Thr	Leu	720
Ser	Ile	Glu	Glu	Asp 725	Ser	Phe	Asn	Gly	Trp 730	Ser	Gln	Ile	Asp	Asp 735	Leu
Ser	Asp	Glu	Asp 740	Asp	Asn	Asp	Gly	Asp 745	Ile	Ala	Ser	Gly	Phe 750	Asn	Phe
Lys	Leu	Ser 755	Thr	Ser	Ala	Pro	Ser 760	Glu	Asn	Val	Asn	Ser 765	His	Thr	Pro
Ile	Leu 770	Gln	Ser	Leu	Asn	Met 775	Ser	Leu	Asp	Gly	A rg 780	Lys	Lys	Xe n	Arg
Ala 785	Ser	Leu	His	Ala	Thr 790	Ser	Val	Leu	Pro	Ser 795	Thr	Ile	Arg	Gln	Nen 800
Asn	Gln	His	Phe	As n 805	Asp	Ile	Asn	Gln	Met 810	Leu	Gly	Ser	Ser	Asp 815	Glu
Asp	Ala	Phe	Pro 820	Lys	Ser	Gln	Ser	Leu 825	Asn	Phe	Asn	Lys	Lys 830	Leu	Pro
Il•	Leu	Lys	11e	Asn	Asp	Asn	Val	11e	Gln	Ser	Asn	Ser	Asn	Ser	Asn

Asn Arg Val Asp Asn Pro lu Asp Thr Val Asp Ser Ser Val Asp Ile 85Õ 855

Thr Ala Phe Tyr Asp Pro Arg Met Ser Ser Asp Ser Lys Phe Asp Trp

Glu Val Ser Lys Asn His Val Asp Pro Ala Ala Tyr Ser Val Asn Val 890 885

Ala Ser Glu Asn Arg Val Leu Asp Asp Phe Lys Lys Ala Phe Arg Glu 900 905

Lys Arg Lys

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2745 base pairs

 - (B) TYPE: nucleic acid (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: coding sequence of CNI

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

λ	TGCTCCAAT	TCAATACAGA	AAATGATACT	GTAGCTCCAG	TGTTTCCCAT	GGAGCAAGAT	60
λ	TAAATGCAG	CACCTGATGC	CGTCCCACTG	GTGCAGACAA	CAACACTACA	AGTCTTTGTA	120
A	AGCTTGCCG	AACCCATAGT	GTTTTTAAAA	GGATTTGAAA	CTAACGGACT	GTCTGAAATA	180
G	CCCCAGTA	TCTTACGAGG	ATCTCTTATC	GTCAGGGTGT	TGAAACCGAA	TAAATTAAAA	240
A	GTATATOGA	TAACCTTCAA	AGGAATATCC	AGAACAGAGT	GGCCGGAAGG	TATACCACCG	300
λ	AGAGAGAAG	AATTTTCAGA	TGTTGAAACT	GTTGTCAATC	ACACATGGCC	ATTTTATCAG	360
G	CGGATGACG	GCATGAATTC	TTTCACCTTA	GAACATCACA	GCTCAAATAA	TTCGTCCAAT	420
C	CCCATCTA	TGAGCGATGA	AGATTATCTA	CTTGAAAAAA	GCGGTGCTTC	AGTATATATC	480
C	CACCAACCG	CTGAACCCCC	TAAAGATAAT	AGCAATCTAA	GTCTGGATGC	CTATGAGCGC	540
M	ACTCATTGT	CATCCGATAA	TTTGAGTAAC	AAGCCAGTAT	CAAGTGATGT	TTCCCATGAC	600
G	ACAGTAAAC	TGTTGGCTAT	TCAAAAGACA	CCATTACCAT	CATCTAGTCG	AAGAGGATCG	660
G?	PACCGGCAA	ATTTTCACGG	TAACTCTTTG	TCACCTCATA	CCTTCATATC	TGATTTGTTC	720
A	CAAAAACAT	TCAGTAATAG	TGGCGCTACT	CCAAGTCCTG	AGCAAGAGGA	TAACTATCTT	780
AC	CACCATCCA	AAGATTCTAA	AGAAGTTTTT	ATTTTTCGAC	CGGGCGATTA	TATTTACACT	840
T	TGAACAGC	CAATATCGCA	ATCTTATCCA	GAAAGTATAA	AAGCCAATTT	TGGTTCCGTG	900
G)	GTATAAAC	TGTCAATAGA	CATAGAGAGG	TTTGGCGCAT	TCAAATCAAC	TATACATACT	960

CAATTACCCA	TCAAAGTCGT	AAGGCTTCCT	TCTGATGGAT	CCGTAGAAGA	GACTGAAGCT	1020
ATTGCAATTT	CCAAGGACTG	GAAAGATCTT	CTTCATTATG	ACGTGGTAAT	TTTCTCGAAA	1080
GAGATCGTTT	TGAATGCATT	TTTACCCATC	GATTTCCATT	TCGCTCCTCT	AGATAAAGTT	1140
ACTCTGCATC	GTATTAGAAT	TTATCTAACA	GAGTCTATGG	AATACACTTG	TANTAGTAAT	1200
GGAAATCACG	AGAAGGCTCG	TAGATTAGAG	CCAACTAAAA	AGTTTCTGTT	GGCTGAACAT	1260
AACGGTCCTA	AACTGCCTCA	TATACCAGCT	GGTTCGAATC	CTTTGAAGGC	TAAAAATAGA	1320
GGGAACATCC	TCTTGGATGA	AAAATCCGGC	GATCTAGTTA	ACAAAGATTT	TCAGTTCGAG	1380
GTGTTTGTCC	CAAGCAAGTT	TACAAACAGT	ATACGGTTAC	ACCCTGATAC	AAATTATGAT	1440
AAAATCAAAG	CCCACCATTG	GATAAAAATT	TGCCTTCGTC	TTTCCAAGAA	GTACGGGGAC	1500
AATAGAAAAC	ATTTCGAAAT	AAGTATTGAT	TCTCCAATCC	ATATTTTAAA	TCAACTATGC	1560
TCACACGCGA	ATACTTTGCT	ACCGAGCTAC	GAGAGTCATT	TCCAGTATTG	TGATGAAGAT	1620
GGTAATTTCG	CACCAGCAGC	AGATCAACAA	AATTACGCAA	GTCATCATGA	TTCCAATATT	1680
TTCTTCCCAA	AAGAAGTTCT	TTCGTCTCCC	GTTCTTTCAC	CTAACGTGCA	GAAGATGAAC	1740
ATTAGAATAC	CGTCTGATCT	TCCAGTAGTG	CGTAATAGAG	CTGAAAGCGT	AAAGAAAAGC	1800
AAGTCAGATA	ATACCTCCAA	GAAGAATGAT	CAAAGTAGCA	ATGTCTTCGC	ATCCAAACAG	1860
CTGGTCGCAA	ACATTTATAA	GCCCAATCAG	ATTCCAAGAG	AATTAACTTC	TCCTCAGGCG	1920
TTACCATTAT	CGCCCATCAC	CTCACCAATT	CTCAATTACC	AACCATTATC	AAACTCCCCG	1980
CCTCCAGATT	TTGATTTTGA	TCTAGCTAAG	CGCGGCGCAG	CCGATTCTCA	TGCTATTCCT	2040
GTGGATCCTC	CATCATATTT	TGATGTATTA	AAGGCCGATG	GGATTGAATT	GCCATACTAC	2100
GATACAAGTT	CATCTAAAAT	TCCTGAACTA	AAACTAAACA	AATCTAGAGA	GACATTGGCC	2160
AGCATTGAGG	AGGACTCATT	CAATGGTTGG	TCTCAAATTG	ATGACTTATC	CGACGAAGAT	2220
GACAATGATG	GCGATATAGC	ATCTGGTTTC	AACTTCAAGC	TGTCAACCAG	TGCTCCGAGT	2280
GAGAACGTTA	ATTCACACAC	TCCTATTTTG	CAGTCTTTAA	ACATGAGTCT	TGATGGGAGA	2340
AAAAAAAATC	GTGCCAGTCT	ACACGCAACA	TCAGTGTTAC	CTAGTACAAT	AAGACAGAAC	2400
AATCAGCATT	TCAATGACAT	AAACCAGATG	CTAGGCAGTA	GTGACGAAGA	TGCCTTTCCC	2460
AAAAGCCAAT	CATTAAATTT	CAATAAGAAA	CTACCAATAC	TTAAAATTAA	TGATAACGTC	2520
АТАСААТСАА	ACAGCAATAG	TAATAACAGA	GTTGATAATC	CAGAAGATAC	AGTGGATTCT	2580
TCAGTCGATA	TTACAGCATT	TTATGATCCA	AGAATGTCAT	CAGATTCCAA	ATTTGATTGG	2640
GAGGTAAGCA	AGAACCATGT	TGACCCAGCA	GCCTACTCGG	TTAACGTTGC	TAGTGAAAAC	2700
CGTGTACTGG	ACGACTTTAA	GAAAGCATTT	CGCGAAAAGA	GAAAA		2745

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single

PCT/US95/13580

51

	31	
	(D) TOPOLOGY: linear	
(ii) l	MOLECULE TYPE: DNA	
(iii) 1	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(vi) (ORIGINAL SOURCE: (C) INDIVIDUAL ISOLATE: CNI-PRC-A	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:7:	
CCAAAAAAA	g agatetegga teaaagtage	30
(2) INFOR	MATION FOR SEQ ID NO:8:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 30 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) 1	MOLECULE TYPE: DNA	
(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(vi)	ORIGINAL SOURCE: (C) INDIVIDUAL ISOLATE: CNI-PCR-B	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:8:	
GGGTTTTTC	A GTGTCGACGA TTCATAGATC	30
(2) INFOR	MATION FOR SEQ ID NO:9:	
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 1964 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: unknown	
(ii)	MOLECULE TYPE: DNA (genomic)	
(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(vi)	ORIGINAL SOURCE: (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full CNA1 coding sequence	
(ix)	FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 2861944	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:9:	

GTTTGTTGCA TTTTGATATT CATCTATATC TATTTCAAAA TTTTTCATGT CATCGCCTCT 60

TGA	AACA:	rga i	ATTT	CCA	AT TO	CTGAI	AAAA	AAG	CGTAC	CTAC	TGG	CAAA	CAA A	AAGG	KAAAS	120
TGT	NTAAJ	ATC (CTTT	\ATG	TT T	TGA	ATCAJ	A GAG	GCAT	TAT	TAT	AAA	GAA (CGAA	CAAAG	180
CCT	CTAA!	TAT	TTGCT	TTA?	KT AJ	AAGG?	(TTA)	Y TTO	CAAA:	AAA	AGTT	CTTT	TTA (GATT	CTTTTT	240
TTT	PTGAG	CGT I	ATTAC	CTCI	AG C	rgcci	LAKTA	A ACI	ACTCI	CAA	CGC			CG Aler Ly		294
			TCT Ser													342
			GAC Amp													390
			ATT													436
			AAG Lys 55													486
GGA Gly	TTA Leu	CCT Pro 70	AAT Asn	CAT His	TCC Ser	TTT Phe	TTA Leu 75	AGA Arg	GAG Glu	CAT His	TTC Phe	TTT Phe 80	CAT His	GAG Glu	GGT Gly	534
			AAG Lys													582
GCA Ala 100	TTG Leu	AGT Ser	AAA Lys	GAA Glu	CCC Pro 105	AAT Asn	CTA Leu	CTA Leu	AAA Lys	CTC Leu 110	AAA Lys	GCG Ala	CCA Pro	ATT Ile	ACT Thr 115	630
ATA Ile	TGT Cys	GGT Gly	GAT Asp	ATT Ile 120	CAC His	GGG Gly	CAG Gln	TAT Tyr	TAT Tyr 125	GAT Asp	TTA Leu	TTG Leu	AAA Lys	CTG Leu 130	TTT Phe	678
GAA Glu	GTT Val	GGC Gly	GGT Gly 135	GAC Asp	CCC Pro	GCC Ala	GAA Glu	ATC Ile 140	GAC Asp	TAT Tyr	TTA Leu	TTC Phe	TTG Leu 145	GCG Gly	GAT Asp	726
TAT Tyr	GTT Val	GAT Asp 150	AGA Arg	GGT Gly	GCA Ala	TTC Phe	TCT Ser 155	TTT Phe	GAG Glu	TGT Cys	CTG Leu	ATT Ile 160	TAT Tyr	TTG Leu	TAC Tyr	774
TCC Ser	TTG Leu 165	AAG Lys	TTG Leu	AAT Asn	AAT Asn	TTA Leu 170	GGG Gly	AGA Arg	TTT Phe	TGG Trp	ATG Met 175	CTA Leu	AGA Arg	GGT Gly	AAC Asn	822
CAT His 180	GAG Glu	TGT Cys	AAG Lys	CAC His	TTG Leu 185	ACC Thr	TCT Ser	TAT Tyr	TTT Phe	ACT Thr 190	TTT Phe	AAG Lys	AAT Asn	GAG Glu	ATG Met 195	870
TTG Leu	CAC His	AAA Lys	TAC Tyr	GAT Asp 200	ATG Met	GAA Glu	GTT Val	TAC Tyr	GAT Asp 205	GCT Ala	TGC Cys	TGC Cys	AGA Arg	TCA Ser 210	TTC Phe	918
AAT Asn	GTC Val	TTA Leu	CCA Pro	TTA Leu	GCA Ala	GCT Ala	TTA Leu	ATG Met	AAC Asn	GGA Gly	CAA Gln	TAT Tyr	TTT Phe	Cys	GTG Val	966

CAT His	G T	GGT Gly 230	Ile	TCT Ser	CCA Pro	AG Glu	TTA Leu 235	Lys	TCA Ser	GTA Val	GAG Glu	GAT Asp 240	Val	AAT Asn	AAA Lys	1014
ATT	AAT Asn 245	Arg	TTT	CGA Arg	GAA Glu	Ile 250	CCA Pro	TCT	CGT	GGT Gly	CTC Leu 255	ATG Met	TGT Cys	GAC Asp	CTA Leu	1062
CTA Leu 260	Trp	GCC Ala	GAT Asp	CCT Pro	GTC Val 265	Glu	AAT Asn	TAT Tyr	GAT Asp	GAT Asp 270	Ala	AGA Arg	GAT Asp	GGT Gly	AGC Ser 275	1110
						GAT Asp									GGT	1158
TGC Cy#	TCT Ser	TTC Phe	GCC Ala 295	TTC Phe	ACT	TTT Phe	AAA Lys	GCA Ala 300	TCA Ser	TGC Cys	AAG Lys	TTT Phe	TTG Leu 305	AAG Lys	GCA Ala	1206
AAT Asn	GGT Gly	TTG Leu 310	TTA Leu	TCT Ser	ATT Ile	ATT Ile	AGA Arg 315	GCA Ala	CAC His	GAA Glu	GCA Ala	CAG Gln 320	GAT Asp	GCT Ala	GGG Gly	1254
TAC Tyr	AGA Arg 325	ATG Met	TAT Tyr	AAA Lys	AAC Asn	AAT Asn 330	AAA Lys	GTA Val	ACA Thr	GGC	TTC Phe 335	CCG Pro	AGC Ser	TTA Leu	ATA Ile	1302
ACC Thr 340	ATG Met	TTC Phe	AGT Ser	GCG Ala	CCA Pro 345	AAC Asn	TAC Tyr	CTG Leu	GAC Asp	ACA Thr 350	TAT Tyr	CAT His	AAT Asn	AAA Lys	GCT Ala 355	1350
GCT Ala	GTG Val	TTA Leu	AAA Lys	TAT Tyr 360	GAA Glu	GAA Glu	AAC Asn	GTC Val	ATG Met 365	AAC Asn	ATC Ile	AGG Arg	CAG Gln	TTT Phe 370	CAC His	1398
ATG Met	TCT Ser	CCG Pro	CAC His 375	CCT Pro	TAC Tyr	TGG Trp	TTG Leu	CCT Pro 380	GAT Asp	TTT Phe	ATG Met	GAT Asp	GTT Val 385	TTC Phe	ACC Thr	1446
TGG	TCA Ser	CTA Leu 390	CCT Pro	TTT Phe	GTT Val	GGC Gly	G AA Glu 395	AAA Lys	GTT Val	ACT Thr	AGC Ser	ATG Met 400	TTA Leu	GTG Val	TCT Ser	1494
ATA Ile	TTA Leu 405	AAC Aen	ATA Ile	TGT Cys	AGT Ser	GAG Glu 410	CAG Gln	GAA Glu	CTT Leu	GAC Asp	CCA Pro 415	GAA Glu	TCG Ser	GAA Glu	CCC Pro	1542
AAA Lys 420	GCT Ala	GCG Ala	GAG Glu	GAG Glu	ACT Thr 425	GTA Val	AAG Lys	GCA Ala	AGA Arg	GCA Ala 430	AAC Asn	GCA Ala	act Thr	AAG Lys	GAG Glu 435	1590
ACC Thr	GGC Gly	ACC Thr	CCA Pro	TCT Ser 440	GAT Asp	G AA Glu	AAG Lys	GCG Ala	TCA Ser 445	TCA Ser	GCG Ala	ATA Ile	TTA Leu	GAA Glu 450	GAT Asp	1638
GAA Glu	ACC Thr	CGA Arg	AGA Arg 455	AAG Lys	GCT Ala	TTG Leu	Arg	AAT Asn 460	AAG Lys	ATA Ile	TTA Leu	GCT Ala	ATT 11= 465	GCT Ala	AAA Lys	1686
GTT Val						Val										1734
TAT Tyr					Asn											1782

CGT Arg 500	GGG Gly	ACT Thr	GAA Glu	GGT Gly	TTG Leu 505	AAT Asn	GAA Glu	ACG Thr	CTA Leu	AGC Ser 510	ACT Thr	TTT Phe	GAA Glu	AAG Lys	GCT Ala 515	1830
AGA Arg	AAG Lys	GAA Glu	GAC Asp	CTT Leu 520	ATT Ile	AAT Asn	GAA Glu	AAA Lys	TTA Leu 525	CCA Pro	CCA Pro	TCT Ser	TTA Leu	TCG Ser 530	GAG Glu	1878
GTT Val	GAA Glu	CAA Gln	GAG Glu 535	AAG Lys	ATT Ile	AAA Lys	TAC Tyr	TAC Tyr 540	GAA Glu	AAA Lys	ATA Ile	TTA Leu	AAG Lys 545	GGA Gly	GCG Ala	1926
			CCA Pro			TGA:	TAAA?	rct :	CAT:	rtt a :	ΓT					1964

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 553 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(vi) SEQUENCE DESCRIPTION: SEO ID NO:10:

	(>	(i) S	EQUE	ENCE	DESC	RIPT	CION:	SEÇ	DID	NO: 1	10:				
Met 1	Ser	Lys	Asp	Leu 5	Asn	Ser	Ser	Arg	Ile 10	Lys	Ile	Ile	Lys	Pro 15	Asn
Asp	Ser	Tyr	11e 20	Lys	Val	Asp	Arg	Lys 25	Lys	Asp	Leu	Thr	Lys 30	Tyr	Glu
Leu	Glu	Asn 35	Gly	Lys	Val	Ile	Ser 40	Thr	Lys	Asp	Arg	Ser 45	Tyr	Ala	Ser
Val	Pro 50	Ala	Ile	Thr	Gly	Lys 55	11e	Pro	Ser	Asp	Glu 60	Glu	Val	Phe	Asp
Ser 65	Lys	Thr	Gly	Leu	Pro 70	Asn	His	Ser	Phe	Leu 75	λrg	Glu	His	Phe	Ph e 80
His	Glu	Gly	λrg	Leu 85	Ser	Lys	Glu	Gln	Ala 90	Ile	Lys	Ile	Leu	Asn 95	Met
Ser	Thr	Val	Ala 100	Leu	Ser	Lys	Glu	Pro 105	Asn	Leu	Leu	Lys	Leu 110	Lys	Ala
Pro	Ile	Thr 115	Ile	Cys	Gly	Asp	Ile 120	His	Gly	Gln	Tyr	Tyr 125	Asp	Leu	Leu
Lys	Leu 130	Phe	Glu	Val	Gly	Gly 135	Asp	Pro	Ala	Glu	11e 140	Asp	Tyr	Leu	Phe
Leu 145	Gly	Asp	Tyr	Val	Asp 150	Arg	GJA	Ala	Phe	Ser 155	Phe	Glu	Cys	Leu	11e 160
Tyr	Leu	Tyr	Ser	Leu 165	Lys	Leu	Asn	Asn	Leu 170	Gly	λrg	Phe	Trp	Met 175	Leu
Arg	Gly	Aen	His 180	Glu	Суя	Lys	Hi=	Leu 185	Thr	Ser	Tyr	Phe	Thr 190	Phe	Lys
Asn	Glu	Met 195	Leu	His	Lys	Tyr	Asp 200	Met	Glu	Val	Tyr	Asp 205	Ala	Cys	Cys

Arg Ser Phe Asn Val Leu Pro Leu Ala Ala Leu Met Asn Gly Gln Tyr Phe Cys Val His Gly Gly Ile Ser Pro Glu Leu Lys Ser Val Glu Asp Val Asn Lys Ile Asn Arg Phe Arg Glu Ile Pro Ser Arg Gly Leu Met Cys Asp Leu Leu Trp Ala Asp Pro Val Glu Asn Tyr Asp Asp Ala Arg Asp Gly Ser Glu Phe Asp Gln Ser Glu Asp Glu Phe Val Pro Asn Ser 280 Leu Arg Gly Cys Ser Phe Ala Phe Thr Phe Lys Ala Ser Cys Lys Phe Leu Lys Ala Asn Gly Leu Leu Ser Ile Ile Arg Ala His Glu Ala Gln Amp Ala Gly Tyr Arg Met Tyr Lym Amn Amn Lym Val Thr Gly Phe Pro 330 Ser Leu Ile Thr Met Phe Ser Ala Pro Asn Tyr Leu Asp Thr Tyr His Asn Lys Ala Ala Val Leu Lys Tyr Glu Glu Asn Val Met Asn Ile Arg Gln Phe His Met Ser Pro His Pro Tyr Trp Leu Pro Asp Phe Met Asp Val Phe Thr Trp Ser Leu Pro Phe Val Gly Glu Lys Val Thr Ser Met Leu Val Ser Ile Leu Asn Ile Cys Ser Glu Gln Glu Leu Asp Pro Glu Ser Glu Pro Lys Ala Ala Glu Glu Thr Val Lys Ala Arg Ala Asn Ala Thr Lys Glu Thr Gly Thr Pro Ser Asp Glu Lys Ala Ser Ser Ala Ile Leu Glu Asp Glu Thr Arg Arg Lys Ala Leu Arg Asn Lys Ile Leu Ala Ile Ala Lys Val Ser Arg Het Phe Ser Val Leu Arg Glu Glu Ser Glu Lys Val Glu Tyr Leu Lys Thr Met Asn Ala Gly Val Leu Pro Arg Gly 490 Ala Leu Ala Arg Gly Thr Glu Gly Leu Asn Glu Thr Leu Ser Thr Phe Glu Lys Ala Arg Lys Glu Asp Leu Ile Asn Glu Lys Leu Pro Pro Ser Leu Ser Glu Val Glu Glu Glu Lys Ile Lys Tyr Tyr Glu Lys Ile Leu 530 540 Lys Gly Ala Glu Lys Lys Pro Gln Leu

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2353 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full CNA2 coding sequence
- (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 262..2073
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

(0.1) 1120101		•	
ATAGTCTATA ATACGTTTGA	PACAGCTAGA TATCGCTAGC	GCCANCATTG TCCCCCTCTC	60
TTGATCAATG CTTTTTTCG	GCCCGAGACA AATGAGAAAA	TGTCCTARAR ATACCTTTCA	120
TCAAGACTCC TATTTTTCCT	PAGAAAAAC ATATATCCAA	CTGGAACAGT ATTAAGCCAA	180
TTGCTACGAT ACAAACAAAA	GGAGATATTC CTTCCCTCCC	ATAGAGTCAC ACAGGAGCCA	240
GTACTTCTTC TTGAACCCGC		ATA AGA AAT ACT GAG Ile Arg Asn Thr Glu 10	291
CAG ATA AAC GCC GCT AT Gln Ile Asn Ala Ala Ile 15	r AAA ATT ATA GAA AAC e Lys Ile Ile Glu Asn 20	AAA ACA GAG CGT CCG Lym Thr Glu Arg Pro 25	339
CAA TCG TCC ACA ACC CC Gln Ser Ser Thr Thr Pro 30			387
GCT AAT TCC ACG GCC ACA Ala Asn Ser Thr Ala The 45			435
CTA GAT GAC GGA AGA GTO Leu Asp Asp Gly Arg Va 60	C GTA TCG ACA AAC CGC 1 Val Ser Thr Asn Arg 65	AGA ATA ATG AAT AAA Arg Ile Met Asn Lys 70	483
GTG CCC GCC ATC ACG TC: Val Pro Ala Ile Thr Se: 75	r His Val Pro Thr Asp	GAA GAG CTG TTC CAG Glu Glu Leu Phe Gln 90	531
CCC AAT GGG ATA CCT CG Pro Asn Gly Ile Pro Arc 95	r CAC GAA TTC CTA AGA g His Glu Phe Leu Arg 100	GAT CAT TTC AAG CGC Asp His Phe Lys Arg 105	579
GAG GGC AAA TTG TCG GC Glu Gly Lys Leu Ser Ala 110	r GC CAG GCG GCC AGG a Ala Gln Ala Ala Arg 115	ATC GTT ACA CTT GCA Ile Val Thr Leu Ala 120	627
ACG GAA CTC TTC AGC AAI Thr Glu Leu Phe Ser Lyn 125	A GAA CCC AAC CTT ATA B Glu Pro Abn Leu Ile 130	TCT GTT CCC GCC CCA Ser Val Pro Ala Pro 135	675

	Val				His			Asp		AAG Lys	723
Phe				Asp						TTG Leu 170	771
			Arg				Phe			TAT Tyr	819
		Lys	CTG Leu			Asp				AGG Arg	867
			AAG Lys							AAT Asn	915
			TAC Tyr								963
			CCC Pro 240								1011
			ATA Ile								1059
			TTC Phe								1107
			GAC Asp								1155
			GAA Glu								1203
			GCA Ala 320								1251
			TAT Tyr								1299
			CTG Leu								1347
			ATG Met								1395
			TTC Phe								1443
			TTG Leu 400								1491

Gln	Phe	AAC	Met	Thr 415	Pro	His	Pro	TAT	Trp 420	Leu	Pro	Asp	Phe	Met 425	Asp	1539
GTT Val	TTC Phe	ACG Thr	TGG Trp 430	TCC Ser	TTG Leu	CCA Pro	TTT Phe	GTT Val 435	GGT Gly	GAA Glu	AAA Lye	GTT Val	ACA Thr 440	GAG Glu	ATG Met	1587
					AAC Asn											1635
															CAA Gln	1683
					GCA Ala 480											1731
AAA Lys	CAT His	GCT Ala	TCC Ser	ATT Ile 495	TTA Leu	GAT Asp	GAC Asp	GAA Glu	CAT His 500	CGA Arg	AGG Arg	AAA Lys	GCC Ala	TTA Leu 505	CGA Arg	1779
					GTC Val											1827
					AAA Lys											1875
					GCT Ala											1923
					GAA Glu 560											1971
AAA Lys	TTA Leu	CCG Pro	CCT Pro	TCA Ser 575	CTA Leu	GAC Amp	GAA Glu	CTG Leu	AAA Lys 580	AAC Asn	GAA Glu	AAT Asn	AAG Lys	AAG Lys 585	TAC Tyr	2019
TAC Tyr	G AA Glu	AAA Lys	GTT Val 590	TGG Trp	CAG Gln	AAA Lys	GTA Val	CAT His 595	GAA Glu	CAT His	GAT Asp	GCA Ala	AAG Lys 600	AAT Asn	GAT Asp	2067
	AAA Lys	TAG	AGAAJ	AGC :	CCT	ATTT(CC AC	CTGT	ACATI	A CTI	CAAT	PAAG	TAAC	STAAC	STT	2123
GCAT	TAAT	TA 7	CTAT	ATTI	SA AC	CTA	SATG	C TC	CTCA	AATG	CAC	AGAA!	rca :	OKTAT	CCTTT	2183
TAT	ragg:	CT (STTC	TTTA!	rt ti	AGT	TTG	r TG	ATCT	CTAT	GAAC	GTA:	TAT :	TAT?	ATGCAA	2243
AAA!	AAA1	CTT 1	LAAT 1	TAT	CT AT	rgga?	CCT	r ac	CAA?	TGT	ATAC	BACG!	CTT ?	rtca?	raggag	2303
TGC	AAAT:	TAT (GAC	ACCA	CC T	CTAI	ATTG	A GC	AGAAC	CGG	TTC	rgaa:	TTC			2353

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 604 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Ser Ser Asp Ala Ile Arg Asn Thr Glu Gln Ile Asn Ala Ala Ile Lys Ile Ile Glu Asn Lys Thr Glu Arg Pro Gln Ser Ser Thr Thr Pro Ile Asp Ser Lys Ala Ser Thr Val Ala Ala Ala Asn Ser Thr Ala Thr Glu Thr Ser Arg Asp Leu Thr Gln Tyr Thr Leu Asp Asp Gly Arg Val Val Ser Thr Asn Arg Arg Ile Met Asn Lys Val Pro Ala Ile Thr Ser His Val Pro Thr Asp Glu Glu Leu Phe Gln Pro Asn Gly Ile Pro Arg His Glu Phe Leu Arg Asp His Phe Lys Arg Glu Gly Lys Leu Ser Ala Ala Gln Ala Ala Arg Ile Val Thr Leu Ala Thr Glu Leu Phe Ser Lys 120 Glu Pro Asn Leu Ile Ser Val Pro Ala Pro Ile Thr Val Cys Gly Asp Ile His Gly Gln Tyr Phe Asp Leu Leu Lys Leu Phe Glu Val Gly Gly Asp Pro Ala Thr Thr Ser Tyr Leu Phe Leu Gly Asp Tyr Val Asp Arg Gly Ser Phe Ser Phe Glu Cys Leu Ile Tyr Leu Tyr Ser Leu Lys Leu Asn Phe Asn Asp His Phe Trp Leu Leu Arg Gly Asn His Glu Cys Lys His Leu Thr Ser Tyr Phe Thr Phe Lys Asn Glu Met Leu His Lys Tyr Asn Leu Asp Ile Tyr Glu Lys Cys Cys Glu Ser Phe Asn Asn Leu Pro Leu Ala Ala Leu Met Asn Gly Gln Tyr Leu Cys Val His Gly Gly Ile 245 250 255 Ser Pro Glu Leu Asn Ser Leu Gln Asp Ile Asn Asn Leu Asn Arg Phe Arg Glu Ile Pro Ser His Gly Leu Met Cys Asp Leu Leu Trp Ala Asp Pro Ile Glu Glu Tyr Asp Glu Val Leu Asp Lys Asp Leu Thr Glu Glu Asp Ile Val Asn Ser Lys Thr Met Val Pro His His Gly Lys Met Ala Pro Ser Arg Asp Met Phe Val Pro Asn Ser Val Arg Gly Cys Ser Tyr 330 325

Ala Phe Thr Tyr Arg Ala Ala Cys His Phe Leu Gln Glu Thr Gly Leu Leu Ser Ile Ile Arg Ala His Glu Ala Gln Asp Ala Gly Tyr Arg Met Tyr Lys Asn Thr Lys Thr Leu Gly Phe Pro Ser Leu Leu Thr Leu Phe Ser Ala Pro Asn Tyr Leu Asp Thr Tyr Asn Asn Lys Ala Ala Ile Leu Lys Tyr Glu Asn Asn Val Met Asn Ile Arg Gln Phe Asn Met Thr Pro His Pro Tyr Trp Leu Pro Asp Phe Het Asp Val Phe Thr Trp Ser Leu Pro Phe Val Gly Glu Lys Val Thr Glu Met Leu Val Ala Ile Leu Asn Ile Cys Thr Glu Asp Glu Leu Glu Asn Asp Thr Pro Val Ile Glu Glu Leu Val Gly Thr Asp Lys Leu Pro Gln Ala Gly Lys Ser Glu Ala 465 470 480 Thr Pro Gln Pro Ala Thr Ser Ala Ser Pro Lys His Ala Ser Ile Leu Asp Asp Glu His Arg Arg Lys Ala Leu Arg Asn Lys Ile Leu Ala Val Ala Lys Val Ser Arg Met Tyr Ser Val Leu Arg Glu Glu Thr Asn Lys Val Gln Phe Leu Lys Asp His Asn Ser Gly Val Leu Pro Arg Gly Ala Leu Ser Asn Gly Val Lys Gly Leu Asp Glu Ala Leu Ser Thr Phe Glu Arg Ala Arg Lys His Asp Leu Ile Asn Glu Lys Leu Pro Pro Ser Leu Asp Glu Leu Lys Asn Glu Asn Lys Lys Tyr Tyr Glu Lys Val Trp Gln Lys Val His Glu His Asp Ala Lys Asn Asp Ser Lys

- (2) INFORMATION FOR SEQ ID NO:13:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 812 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double (D) TOPOLOGY: unknown
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO
 - (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: genomic DNA fragment containing full

CNB1 coding sequence

(ix) FEATURE:

(A) NAME/KEY: CDS (B) LOCATION: 54..104

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 181..652

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

ACTTGGTAAC TCAATGGTGA TCAGAATCCA TAGAAGCATT TTTATTTCTT AAA ATG Met 1	56
GGT GCT GCT CCT TCC AAA ATT GTG GAT GGT CTT TTA GAA GAT ACA AAT Gly Ala Ala Pro Ser Lys Ile Val Asp Gly Leu Leu Glu Asp Thr Asn 5 10 15	104
TGTATGTACA CTTCGGAGTG AGGAAAAGAA AGAAAGGGGA AATTAACCGA TTTTACTAAC	164
ACTGACACTT TGAACA GTT GAT AGA GAT GAA ATT GAA AGG TTA AGG AAG Val Asp Arg Asp Glu Ile Glu Arg Leu Arg Lys 1 5 10	213
AGA TTC ATG AAA TTA GAT AGA GAT AGC TCA GGG TCT ATT GAT AAA AAT Arg Phe Met Lys Leu Asp Arg Asp Ser Ser Gly Ser Ile Asp Lys Asn 15 20 25	261
GAA TTT ATG AGC ATT CCT GGC GTT TCG TCA AAC CCT CTT GCT GGA CGT Glu Phe Met Ser Ile Pro Gly Val Ser Ser Asn Pro Leu Ala Gly Arg 30 35 40	309
ATA ATG GAG GTT TTC GAT GCT GAT AAT AGT GGG GAC GTG GAT TTT CAA Ile Het Glu Val Phe Asp Ala Asp Asn Ser Gly Asp Val Asp Phe Gln 45 50 55	357
GAG TTC ATC ACA GGA TTA TCC ATT TTC AGT GGG CGT GGG TCC AAG GAC Glu Phe Ile Thr Gly Leu Ser Ile Phe Ser Gly Arg Gly Ser Lys Asp 60 65 70 75	405
GAA AAG TTA AGA TTC GCC TTC AAA ATC TAC GAC ATT GAC AAG GAC GGT Glu Lys Leu Arg Phe Ala Phe Lys Ile Tyr Asp Ile Asp Lys Asp Gly 80 85 90	453
TTC ATA TCC AAT GGT GAG TTG TTC ATC GTG TTG AAG ATT ATG GTA GGT Phe Ile Ser Asn Gly Glu Leu Phe Ile Val Leu Lys Ile Met Val Gly 95 100 105	501
TCT AAT CTG GAC GAT GAA CAG CTG CAA CAG ATA GTA GAT AGG ACG ATA Ser Asn Leu Asp Asp Glu Gln Leu Gln Gln Ile Val Asp Arg Thr Ile 110 115 120	549
GTG GAA AAC GAT AGC GAC GGC GAC GGA CGT TTA AGT TTC GAG GAG TTT Val Glu Asn Asp Ser Asp Gly Asp Gly Arg Leu Ser Phe Glu Glu Phe 125 130 135	5 9 7
AAG AAT GCT ATC GAA ACC ACA GAA GTG GCC AAG AGT CTG ACA TTG CAA Lys Asn Ala Ile Glu Thr Thr lu Val Ala Lys Ser Leu Thr Leu Gln 140 155 155	645
TAC GATGTGTAAG ACTAGGGGAC ACTTCATTCA TTTATGGTAT GCCAATATTT Tyr Amp	698

TTAAGAAAA AAGAATAATA CGCGATATTG TTTTTTAAGG AAGGAACGCA CACTCGCCCA 758
GTTAGAGTGC TGATGATATA TACATATATA TATGTATATG TAACAAACAA TAAG 812

- (2) INFORMATION FOR SEQ ID NO:14:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 amino acids
 - (B) TYPE: amino acid
 (D) TOPOLOGY: linear
 - (5, 101020011 22......
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Gly Ala Ala Pro Ser Lys Ile Val Asp Gly Leu Leu Glu Asp Thr
1 10 15

Asn

- (2) INFORMATION FOR SEQ ID NO:15:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 157 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Val Asp Arg Asp Glu Ile Glu Arg Leu Arg Lys Arg Phe Met Lys Leu
1 1 15

Asp Arg Asp Ser Ser Gly Ser Ile Asp Lys Asn Glu Phe Met Ser Ile 20 25 30

Pro Gly Val Ser Ser Asn Pro Leu Ala Gly Arg Ile Met Glu Val Phe 35 40 45

Asp Ala Asp Asn Ser Gly Asp Val Asp Phe Gln Glu Phe Ile Thr Gly 50 55 60

Leu Ser Ile Phe Ser Gly Arg Gly Ser Lys Asp Glu Lys Leu Arg Phe 65 70 75 80

Ala Phe Lys Ile Tyr Asp Ile Asp Lys Asp Gly Phe Ile Ser Asn Gly 85 90 95

Glu Leu Phe Ile Val Leu Lys Ile Met Val Gly Ser Asn Leu Asp Asp 100 105 110

Glu Gln Leu Gln Gln Ile Val Asp Arg Thr Ile Val Glu Asn Asp Ser 115 120 125

Asp Gly Asp Gly Arg Leu Ser Phe Glu Glu Phe Lys Asn Ala Ile Glu 130 135 140

Thr Thr Glu Val Ala Lys Ser Leu Thr Leu Gln Tyr Asp 145 150 155

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 524 base pairs

 - (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE: (C) INDIVIDUAL ISOLATE: coding sequence of CNB1
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..524

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

ATG Met 1	GGT Gly	GCT Ala	GCT Ala	CCT Pro 5	TCC Ser	AAA Lys	ATT Ile	GTG Val	GAT Asp 10	GGT Gly	CTT Leu	TTA Leu	G AA Glu	GAT Asp 15	ACA Thr	48
AAT ABD	TTT Phe	GAT Asp	AGA Arg 20	GAT Asp	GAA Glu	ATT 11e	GAA Glu	AGG Arg 25	TTA Leu	AGG Arg	AAG Lys	AGA Arg	TTC Phe 30	ATG Met	AAA Lys	96
TTA Leu	GAT Amp	AGA Arg 35	GAT Asp	AGC Ser	TCA Ser	GGG Gly	TCT Ser 40	ATT Ile	GAT Asp	AAA Lys	AAT Asn	GAA Glu 45	TTT Phe	ATG Met	AGC Ser	144
ATT Ile	CCT Pro 50	GGC Gly	GTT Val	TCG Ser	TCA Ser	AAC Aen 55	CCT Pro	CTT Leu	GCT Ala	GGA Gly	CGT Arg 60	ATA Ile	ATG Met	GAG Glu	GTT Val	192
TTC Phe 65	GAT Asp	GCT Ala	GAT Asp	AAT Asn	AGT Ser 70	GGG Gly	GAC Asp	GTG Val	GAT Asp	TTT Phe 75	CAA Gln	GAG Glu	TTC Phe	ATC Ile	ACA Thr 80	240
GGA Gly	TTA Leu	TCC Ser	ATT Ile	TTC Phe 85	AGT Ser	GGG Gly	CGT Arg	GCG	TCC Ser 90	AAG Lys	GAC Asp	GAA Glu	AAG Lys	TTA Leu 95	AGA Arg	288
TTC Phe	GCC Ala	TTC Phe	AAA Lys 100	ATC Ile	TAC Tyr	GAC Asp	ATT	GAC Asp 105	AAG Lys	GAC Asp	GGT Gly	TTC Phe	ATA Ile 110	TCC Ser	AAT Asn	336
GGT Gly	GAG Glu	TTG Leu 115	TTC Phe	ATC Ile	GTG Val	TTG Leu	AAG Lys 120	ATT Ile	ATG Met	GTA Val	GGT Gly	TCT ser 125	AAT Asn	CTG Leu	GAC Asp	384
GAT A ≠ P	GAA Glu 130	CAG Gln	CTG Leu	CAA Gln	CAG Gln	ATA 11e 135	GTA Val	GAT Amp	AGG Arg	ACG Thr	ATA Ile 140	GTG Val	GAA Glu	AAC Asn	GAT Asp	432
AGC Ser 145	GAC Asp	GCC	GAC Amp	GGA Gly	CGT Arg 150	TTA Leu	AGT Ser	TTC Phe	GAG Glu	GAG Glu 155	TTT Phe	AAG Lys	AAT Asn	GCT Ala	ATC Ile 160	480
GAA Glu	ACC Thr	ACA Thr	GAA Glu	GTG Val 165	GCC Ala	AAG Lys	AGT Ser	CTG Leu	ACA Thr 170	TTG Leu	CAA Gln	TAC Tyr	GAT Asp	GT		524

(2) INFORMATION FOR SEQ ID N :17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 174 amino acids

 - (B) TYPE: amino acid (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Met Gly Ala Ala Pro Ser Lys Ile Val Asp Gly Leu Leu Glu Asp Thr

Asn Phe Asp Arg Asp Glu Ile Glu Arg Leu Arg Lys Arg Phe Met Lys

Leu Asp Arg Asp Ser Ser Gly Ser Ile Asp Lys Asn Glu Phe Met Ser

Ile Pro Gly Val Ser Ser Asn Pro Leu Ala Gly Arg Ile Met Glu Val

Phe Asp Ala Asp Asn Ser Gly Asp Val Asp Phe Gln Glu Phe Ile Thr

Gly Leu Ser Ile Phe Ser Gly Arg Gly Ser Lys Asp Glu Lys Leu Arg

Phe Ala Phe Lys Ile Tyr Asp Ile Asp Lys Asp Gly Phe Ile Ser Asn

Gly Glu Leu Phe Ile Val Leu Lys Ile Met Val Gly Ser Asn Leu Asp

Asp Glu Gln Leu Gln Gln Ile Val Asp Arg Thr Ile Val Glu Asn Asp

Ser Asp Gly Asp Gly Arg Leu Ser Phe Glu Glu Phe Lys Asn Ala Ile

Glu Thr Thr Glu Val Ala Lys Ser Leu Thr Leu Gln Tyr Asp

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1812 base pairs
 - (B) TYPE: nucleic acid (C) STRANDEDNESS: double

 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: DNA fragment containing CNAldeltaC coding sequence
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 286..1812

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:																
GTTTGTTGCA TTTTGATATT CATCTATATC TATTTCAAAA TTTTTCATGT CATCGCCTCT												60				
TGAAACATGA ATTTTCCAAT TCTGAAAAAG AACGTACTAC TGGGAAACAA AAGGGAAAAA											120					
TGTATAAATC CTTTAATGTT TTTGAATCAA GAGGCATTAT TATAAAAGAA CGAAGCAAAG										180						
CCTTTARTAT TTGCTTTATT ARAGGTATTA TTCRARGARA AGTTTTTTTA GATTCTTTTT										240						
TTT:	rtga(CGT :	ATTA(CTC	AG C	rgcci	ATAA	A AC	ACTC	TCAA	CGC	CA A'	TG TG	CG A	AA ys	294
GAC Amp	TTG Leu 5	AAT Asn	TCT Ser	TCA Ser	CGC Arg	ATC Ile 10	AAA Lys	ATC Ile	ATT Ile	AAA Lys	CCT Pro 15	AAT Asn	GAC Asp	TCT Ser	TAC Tyr	342
ATA Ile 20	AAG Lys	GTT Val	GAC Asp	CGG Arg	AAA Lys 25	AAA Lys	GAT Asp	TTA Leu	ACA Thr	AAA Lys 30	TAC Tyr	GAA Glu	TTA Leu	GAA Glu	AAC Aan 35	390
GCT Gly	AAA Lys	GTA Val	ATT Ile	TCT Ser 40	ACT Thr	AAG Lys	GAC Asp	CGA Arg	TCC Ser 45	TAC Tyr	GCT Ala	TCT Ser	GTA Val	CCT Pro 50	GCC Ala	438
ATA Ile	ACA Thr	GGA Gly	AAG Lys 55	ATA Ile	CCA Pro	AGT Ser	GAT Asp	GAG Glu 60	GAA Glu	GTA Val	TTC Phe	GAC Asp	TCC Ser 65	AAG Lys	ACG Thr	486
GGA Gly	TTA Leu	CCT Pro 70	Asn	CAT His	TCC Ser	TTT Phe	TTA Leu 75	AGA Arg	GAG Glu	CAT	TTC Phe	TTT Phe 80	CAT His	GAG Glu	GGT Gly	534
CGA Arg	CTT Leu 85	TCT Ser	AAG Lys	GAA Glu	CAG Gln	GCC Ala 90	ATA Ile	AAA Lys	ATC Ile	TTA Leu	AAT Asn 95	ATG Met	TCA Ser	ACT Thr	GTA Val	582
GCA Ala 100	Leu	AGT Ser	λλλ Lys	G AA Glu	CCC Pro 105	AAT Asn	CTA Leu	CTA Leu	AAA Lys	CTC Leu 110	AAA Lys	GCG Ala	CCA Pro	ATT	ACT Thr 115	630
ATA Ile	TGT Cys	GGT Gly	GAT Asp	ATT Ile 120	CAC Him	GGG Gly	CAG Gln	TAT Tyr	TAT Tyr 125	GAT Asp	TTA Leu	TTG Leu	AAA Lys	CTG Leu 130	TTT Phe	678
GAA Glu	GTT Val	GGC Gly	GGT Gly 135	GAC Asp	CCC Pro	GCC Ala	GAA Glu	ATC Ile 140	GAC Amp	TAT Tyr	TTA Leu	TTC Phe	TTG Leu 145	GGG Gly	GAT Asp	726
TAT Tyr	GTT Val	GAT Asp 150	AGA Arg	GGT Gly	GCA Ala	TTC Phe	TCT Ser 155	TTT Phe	GAG Glu	TGT Cys	CTG Leu	Ile 160	TAT	TTG Leu	TAC Tyr	774
TCC Ser	TTG Leu 165	AAG Lys	TTG Leu	AAT Asn	AAT Asn	TTA Leu 170	GGG Gly	AGA Arg	TTT Phe	TGG Trp	ATG Met 175	CTA Leu	AGA Arg	GGT Gly	AAC Asn	822
CAT His 180	GAG Glu	TGT Cys	AAG Lys	CAC His	TTG Leu 185	ACC Thr	TCT Ser	TAT Tyr	TTT Phe	ACT Thr 190	TTT Phe	λλG Ly∎	AAT Asn	GAG Glu	ATG Met 195	870
TTG Leu	CAC His	AAA Lys	TAC Tyr	GAT Asp 200	ATG Met	AA Glu	GTT Val	TAC Tyr	GAT Asp 205	GCT Ala	TGC Cys	TGC Cys	AGA Arg	TCA Ser 210	TTC Phe	918

AAT Asn	GTC Val	TTA Leu	CCA Pro 215	TTA Leu	GCA Ala	GCT Ala	TTA Leu	ATG Met 220	AAC Asn	GGA Gly	CAA Gln	TAT Tyr	TTT Phe 225	TGT Cys	GTG Val	966
CAT His	GGT Gly	GGT Gly 230	ATC Ile	TCT Ser	CCA Pro	GAG Glu	TTA Leu 235	AAA Lys	TCA Ser	GTA Val	GAG Glu	GAT Asp 240	GTT Val	AAT Asn	AAA Lys	1014
					GAA Glu											1062
CTA Leu 260	TGG Trp	GCC Ala	GAT Asp	CCT Pro	GTC Val 265	GAA Glu	AAT Asn	TAT Tyr	GAT Asp	GAT Asp 270	GCA Ala	AGA Arg	GAT Asp	GCT Gly	AGC Ser 275	1110
GAA Glu	TTT Phe	GAT Asp	CAG Gln	AGC Ser 280	GAG Glu	GAT Asp	GAA Glu	TTC Phe	GTA Val 285	CCT Pro	AAC Asn	AGT Ser	TTG Leu	AGG Arg 290	GLY	1158
TGC Cys	TCT Ser	TTC Phe	GCC Ala 295	TTC Phe	ACT Thr	TTT Phe	AAA Lys	GCA Ala 300	TCA Ser	TGC Cys	AAG Lys	TTT Phe	TTG Leu 305	AAG Lys	GCA Ala	1206
AAT Asn	GGT Gly	TTG Leu 310	TTA Leu	TCT Ser	ATT Ile	ATT Ile	AGA Arg 315	GCA Ala	CAC His	GAA Glu	GCA Ala	CAG Gln 320	GAT Asp	GCT Ala	GGG	1254
TAC Tyr	AGA Arg 325	ATG Met	TAT Tyr	AAA Lys	AAC Asn	AAT Asn 330	AAA Lys	GTA Val	ACA Thr	GGC Gly	TTC Phe 335	CCG Pro	AGC Ser	TTA Leu	ATA Ile	1302
					CCA Pro 345											1350
GCT Ala	GTG Val	TTA Leu	AAA Lys	TAT Tyr 360	GAA Glu	GAA Glu	AAC Asn	GTC Val	ATG Met 365	AAC Asn	ATC Ile	AGG Arg	CAG Gln	TTT Phe 370	CAC His	1398
ATG Met	TCT Ser	CCG Pro	CAC His 375	CCT Pro	TAC Tyr	TGG Trp	TTG Leu	CCT Pro 380	GAT Asp	TTT Phe	ATG Met	GAT Asp	GTT Val 385	TTC Phe	ACC Thr	1446
TGG Trp	TCA Ser	CTA Leu 390	CCT Pro	TTT Phe	GTT Val	GGC Gly	GAA Glu 395	AAA Lys	GTT Val	ACT Thr	AGC Ser	ATG Met 400	TTA Leu	GTG Val	TCT Ser	1494
ATA Ile	TTA Leu 405	AAC Asn	ATA Ile	TGT Cys	AGT Ser	GAG Glu 410	CAG Gln	GAA Glu	CTT Leu	GAC Asp	CCA Pro 415	GAA Glu	TCG Ser	GAA Glu	CCC Pro	1542
AAA Lys 420	GCT Ala	GCG Ala	GAG Glu	GAG Glu	ACT Thr 425	GTA Val	AAG Lys	GCA Ala	AGA Arg	GCA Ala 430	AAC Asn	GCA Ala	ACT Thr	AAG Lys	GAG Glu 435	1590
ACC Thr	GGC Gly	ACC Thr	CCA Pro	TCT Ser 440	GAT Asp	GAA Glu	AAG Lys	GCG Ala	TCA Ser 445	TCA Ser	GCG Ala	ATA Ile	TTA Leu	GAA Glu 450	GAT Amp	1638
GAA Glu	ACC Thr	CGA Arg	AGA Arg 455	AAG Lys	GCT Ala	TTG Leu	AGA Arg	AAT Asn 460	AAG Lys	ATA Ile	TTA Leu	GCT Ala	ATT Ile 465	GCT Ala	AAA Lys	1686
GTT Val	TCA Ser	AGA Arg 470	ATG Met	TTT Phe	TCG Ser	GTG Val	CTA Leu 475	AGA Arg	GAA Glu	GAG lu	AGC Ser	GAA Glu 480	AAA Lys	GTG Val	GAA Glu	1734

TAT TTG AAA ACT ATG AAT GCC GGT GTC TTA CCT CGT GGT GCT CTA GCT
TYR Leu Lys Thr Met Asn Ala Gly Val Leu Pro Arg Gly Ala Leu Ala
485

CGT GGG ACT GAA GGT TTG AAT GAA ACG CTA
Arg Gly Thr Glu Gly Leu Asn Glu Thr Leu
500

1782

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 509 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Met Ser Lys Asp Leu Asn Ser Ser Arg Ile Lys Ile Ile Lys Pro Asn Asp Ser Tyr Ile Lys Val Asp Arg Lys Lys Asp Leu Thr Lys Tyr Glu 20 25 30 Leu Glu Asn Gly Lys Val Ile Ser Thr Lys Asp Arg Ser Tyr Ala Ser Val Pro Ala Ile Thr Gly Lys Ile Pro Ser Asp Glu Glu Val Phe Asp Ser Lys Thr Gly Leu Pro Asn His Ser Phe Leu Arg Glu His Phe Phe His Glu Gly Arg Leu Ser Lys Glu Gln Ala Ile Lys Ile Leu Asn Met Ser Thr Val Ala Leu Ser Lys Glu Pro Asn Leu Leu Lys Leu Lys Ala Pro Ile Thr Ile Cys Gly Asp Ile His Gly Gln Tyr Tyr Asp Leu Leu Lys Leu Phe Glu Val Gly Gly Asp Pro Ala Glu Ile Asp Tyr Leu Phe Leu Gly Asp Tyr Val Asp Arg Gly Ala Phe Ser Phe Glu Cys Leu Ile Tyr Leu Tyr Ser Leu Lys Leu Asn Asn Leu Gly Arg Phe Trp Met Leu Arg Gly Asn His Glu Cys Lys His Leu Thr Ser Tyr Phe Thr Phe Lys Asn Glu Het Leu His Lys Tyr Asp Met Glu Val Tyr Asp Ala Cys Cys 200 Arg Ser Phe Asn Val Leu Pro Leu Ala Ala Leu Met Asn Gly Gln Tyr Phe Cys Val His Gly Gly Ile Ser Pro Glu Leu Lys Ser Val Glu Asp Val Asn Lys Ile Asn Arg Phe Arg lu Ile Pro Ser Arg Gly Leu Met 245

- Cys
 Asp
 Leu
 Leu 260
 Trp
 Ala
 Asp
 Pro
 Val 265
 Glu
 Asp
 Asp
 Arg
 Glu
 Asp
 Glu
 Pro
 Asp
 Arg
 Glu
 Asp
 Glu
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 Asp
 Pro
 Asp
 Glu
 Asp
 Glu
 Pro
 Asp
 Pro
 Asp
 Asp
 Glu
 Asp
 <th
- Ile Ala Lys Val Ser Arg Met Phe Ser Val Leu Arg Glu Glu Ser Glu 465 470 475 480

Leu Glu Asp Glu Thr Arg Arg Lys Ala Leu Arg Asn Lys Ile Leu Ala

Thr Lys Glu Thr Gly Thr Pro Ser Asp Glu Lys Ala Ser Ser Ala Ile

Lys Val Glu Tyr Leu Lys Thr Het Asn Ala Gly Val Leu Pro Arg Gly
485 490 495

Ala Leu Ala Arg Gly Thr Glu Gly Leu Asn Glu Thr Leu 500 505

(2) INFORMATION FOR SEQ ID NO:20:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1767 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: DNA fragment containing CNA2deltaC coding sequence
- (ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 262..1767

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

ATAGTCTATA ATAC	GTTTGA TACAG	CTAGA TATCGCTA	AGC GCCAACATTG T	CCCCCTCTC 60
TTGATCAATG CTTI	TTTTCG GCCCG	AGACA AATGAGAI	AAA TGTCCTAAAA A	TACCTTTCA 120
TCAAGACTCC TATI	TTTCCT TAGAA	AAAAC ATATATCO	CAA CTGGAACAGT A	TTAAGCCAA 180
TTGCTACGAT ACAN	ACAAAA GGAGA:	PATTC CTTCCCTC	CC ATAGAGTCAC A	CAGGAGCCA 240
GTACTTCTTC TTGA	ACCCGC A ATG Met 1	TCT TCA GAC C Ser Ser Asp A	GCT ATA AGA AAT Ala Ile Arg Asn 5	ACT GAG 291 Thr Glu 10
CAG ATA AAC GCC Gln Ile Asn Ala	GCT ATT AAA Ala Ile Lys 15	ATT ATA GAA 2 Ile Ile Glu 2 20	AAC AAA ACA GAG Asn Lys Thr Glu	CGT CCG 339 Arg Pro 25
CAA TCG TCC ACA Gln Ser Ser Thr 30	Thr Pro Ile	GAT TCG AAG C Asp Ser Lys A 35	GCT AGT ACA GTT Ala Ser Thr Val 40	GCT GCT 387 Ala Ala
GCT AAT TCC ACG Ala Amn Ser Thr 45	GCC ACA GAA Ala Thr Glu	ACT TCC AGA C Thr Ser Arg 2 50	GAC CTT ACA CAA Asp Leu Thr Gln 55	TAT ACC 435 Tyr Thr
CTA GAT GAC GGA Leu Asp Asp Gly 60	AGA GTC GTA Arg Val Val 65	TCG ACA AAC G Ser Thr Asn A	CGC AGA ATA ATG Arg Arg Ile Met 70	ART ARA 483 Asn Lys
GTG CCC GCC ATC Val Pro Ala Ile 75	ACG TCA CAT Thr Ser His 80	GTT CCT ACA (GAT GAA GAG CTG Asp Glu Glu Leu 85	TTC CAG 531 Phe Gln 90
CCC AAT GGG ATA	CCT CGT CAC Pro Arg His 95	GAA TTC CTA A Glu Phe Leu A 100	AGA GAT CAT TTC Arg Asp His Phe	AAG CGC 579 Lys Arg 105
GAG GGC AAA TTG Glu Gly Lys Leu 110	Ser Ala Ala	CAG GCG GCC I Gln Ala Ala I 115	AGG ATC GTT ACA Arg Ile Val Thr 120	CTT GCA 627 Leu Ala
ACG GAA CTC TTC Thr Glu Leu Phe 125	AGC AAA GAA Ser Lys Glu	CCC AAC CTT I Pro Asn Leu 1 130	ATA TCT GTT CCC ile ser Val Pro 135	GCC CCA 675 Ala Pro
ATC ACA GTT TGC Ile Thr Val Cys 140	GGT GAT ATC Gly Amp Ile 145	CAT GGC CAG	TAC TTT GAC CTT Tyr Phe Asp Leu 150	TTG AAG 723 Leu Lys
CTA TTC GAA GTT Leu Phe Glu Val 155	GGC GGA GAT Gly Gly Amp 160	Pro Ala Thr	ACA TCG TAT TTG Thr Ser Tyr Leu 165	TTC TTG 771 Phe Leu 170
GGA GAC TAT GTG Gly Asp Tyr Val	GAC AGA GGG Amp Arg Gly 175	TCC TTT TCG Ser Phe Ser 180	TTT GAG TGT CTT Phe Glu Cys Leu	ATT TAT 819 Ile Tyr 185
TTA TAT TCT TTC Leu Tyr Ser Leu 190	Lys Leu Asn	TTT AAC GAC (Phe Asn Asp 1	CAT TTC TGG CTA His Phe Trp Leu 200	CTG AGG 867 Leu Arg

GGT Gly	AAC Asn	CAC His 205	GAA Glu	TGT Cys	AAG Lys	CAT His	CTA Leu 210	ACG Thr	TCA Ser	TAT Tyr	TTC Phe	ACT Thr 215	TTC Phe	AAA Lys	AAT Asn	915
					TAC Tyr											963
					CCC Pro 240											1011
					ATA Ile											1059
AAC Asn	AAC Asn	CTA Leu	AAT Asn 270	AGA Arg	TTC Phe	AGG Arg	GAG Glu	ATT Ile 275	CCC Pro	TCT Ser	CAT His	GGC Gly	CTG Leu 280	ATG Met	TGT Cys	1107
					GAC Asp											1155
					GAA Glu											1203
					GCA Ala 320											1251
					TAT Tyr											1299
					CTG Leu											1347
GAC Asp	GCT Ala	GGT Gly 365	TAT Tyr	AGA Arg	ATG Met	TAC Tyr	AAA Lys 370	AAT Asn	ACC Thr	AAG Lys	ACT Thr	TTG Leu 375	GGC Gly	TTT Phe	CCC Pro	1395
TCT Ser	CTT Leu 380	TTG Leu	ACC Thr	CTT Leu	TTC Phe	AGT Ser 385	GCG Ala	CCT Pro	AAC Asn	TAC Tyr	TTG Leu 390	GAC Asp	ACC Thr	TAC Tyr	AAT Asn	1443
AAT Asn 395	AAG Lys	GCT Ala	GCC Ala	ATA Ile	TTG Leu 400	AAA Lys	TAC Tyr	GAA Glu	AAT Asn	AAT Asn 405	GTT Val	ATG Met	AAT Asn	ATC Ile	AGA Arg 410	1491
CAA Gln	TTC Phe	AAC Asn	ATG Met	ACT Thr 415	CCA Pro	CAC His	CCC Pro	TAT Tyr	TGG Trp 420	TTA Leu	CCA Pro	GAT Asp	TTC Phe	ATG Met 425	GAC Asp	1539
GTT Val	TTC Phe	ACG Thr	TGG Trp 430	TCC Ser	TTG Leu	CCA Pro	TTT Phe	GTT Val 435	GGT Gly	GAA Glu	AAA Lys	GTT Val	ACA Thr 440	GAG Glu	ATG Met	1587
CTT Leu	GTC Val	GCA Ala 445	ATT Ile	CTA Leu	AAC Asn	ATC Ile	TGT Cys 450	ACT Thr	G AA Glu	GAT Asp	GAG Glu	CTG Leu 455	GAA Glu	AAC Asn	GAC Asp	1635
ACC Thr	CCC Pro 460	GTC Val	ATT Ile	GAA Glu	GAA Glu	TTA Leu 465	GTT Val	GGT Gly	ACC Thr	GAT Asp	AAA Lys 470	AAA Lys	TT Leu	CCA Pro	CAA Gln	1683

71

GCT GGT ANG TCG GAN GCN ACT CCN CAN CCN GCC ACT TCG GCG TCG CCT 1731 Ala Gly Lys Ser Glu Ala Thr Pro In Pro Ala Thr Ser Ala Ser Pro 485 1767 AAA CAT GCT TCC ATT TTA GAT GAC GAA CAT CGA AGG Lys His Ala Ser Ile Leu Asp Asp Glu His Arg Arg 500 495

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 502 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Met Ser Ser Asp Ala Ile Arg Asn Thr Glu Gln Ile Asn Ala Ala Ile Lys Ile Ile Glu Asn Lys Thr Glu Arg Pro Gln Ser Ser Thr Thr Pro lle Asp Ser Lys Ala Ser Thr Val Ala Ala Ala Asn Ser Thr Ala Thr Glu Thr Ser Arg Asp Leu Thr Gln Tyr Thr Leu Asp Asp Gly Arg Val Val Ser Thr Asn Arg Arg Ile Met Asn Lys Val Pro Ala Ile Thr Ser His Val Pro Thr Asp Glu Glu Leu Phe Gln Pro Asn Gly Ile Pro Arg His Glu Phe Leu Arg Asp His Phe Lys Arg Glu Gly Lys Leu Ser Ala Ala Gln Ala Ala Arg Ile Val Thr Leu Ala Thr Glu Leu Phe Ser Lys Glu Pro Asn Leu Ile Ser Val Pro Ala Pro Ile Thr Val Cys Gly Asp Ile His Gly Gln Tyr Phe Asp Leu Leu Lys Leu Phe Glu Val Gly Gly Asp Pro Ala Thr Thr Ser Tyr Leu Phe Leu Gly Asp Tyr Val Asp Arg Gly Ser Phe Ser Phe Glu Cys Leu Ile Tyr Leu Tyr Ser Leu Lys Leu Asn Phe Asn Asp His Phs Trp Leu Leu Arg Gly Asn His Glu Cys Lys His Leu Thr Ser Tyr Phe Thr Phs Lys Asn Glu Met Leu His Lys Tyr 215 Asn Leu Asp Ile Tyr Glu Lys Cys Cys Glu Ser Phe Asn Asn Leu Pro Leu Ala Ala Leu Het Asn Gly Gln Tyr Leu Cys Val His Gly Gly Ile 250 245

72

Ser Pro Glu Leu Asn Ser Leu Gln Asp Ile Asn Asn Leu Asn Arg Phe 260 265 Arg Glu Ile Pro Ser His Gly Leu Met Cys Asp Leu Leu Trp Ala Asp Pro Ile Glu Glu Tyr Asp Glu Val Leu Asp Lys Asp Leu Thr Glu Glu Asp Ile Val Asn Ser Lys Thr Met Val Pro His His Gly Lys Met Ala 305 320 310 Pro Ser Arg Asp Met Phe Val Pro Asn Ser Val Arg Gly Cys Ser Tyr Ala Phe Thr Tyr Arg Ala Ala Cys His Phe Leu Gln Glu Thr Gly Leu Leu Ser Ile Ile Arg Ala His Glu Ala Gln Asp Ala Gly Tyr Arg Met Tyr Lys Asn Thr Lys Thr Leu Gly Phe Pro Ser Leu Leu Thr Leu Phe Ser Ala Pro Asn Tyr Leu Asp Thr Tyr Asn Asn Lys Ala Ala Ile Leu Lys Tyr Glu Asn Asn Val Met Asn Ile Arg Gln Phe Asn Met Thr Pro 405 His Pro Tyr Trp Leu Pro Asp Phe Met Asp Val Phe Thr Trp Ser Leu Pro Phe Val Gly Glu Lys Val Thr Glu Het Leu Val Ala Ile Leu Asn Cys Thr Glu Asp Glu Leu Glu Asn Asp Thr Pro Val Ile Glu Glu 450 455 Leu Val Gly Thr Asp Lys Leu Pro Gln Ala Gly Lys Ser Glu Ala Thr Pro Gln Pro Ala Thr Ser Ala Ser Pro Lys His Ala Ser Ile Leu 490 Asp Asp Glu His Arg Arg 500

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (C) INDIVIDUAL ISOLATE: G4-PCR-A

2
2

IT IS CLAIMED:

- A polypeptide composition comprising a polypeptide effective to enhance immunosuppressive effects of a calcineurin-targeted immunosuppressant by potentiating an interaction of an immunophilin with calcineurin.
 - 2. A composition of claim 1, wherein the polypeptide composition contains a calcineurin interacting (CNI) polypeptide.
- 3. A composition of claim 2, wherein the polypeptide composition contains a polypeptide having an amino acid sequence selected from the group consisting of sequences represented by SEQ ID NO:2 and SEQ ID NO:5.
- 4. A composition of claim 2, wherein the polypeptide has an amino acid sequence f
 between 15 and 915 amino acids in length.
 - 5. A composition of claim 2, wherein the polypeptide composition contains a polypeptide having an amino acid sequence comprising the c-terminal 306 amino acids of a CNI protein.

20

- 6. An isolated nucleic acid having a sequence encoding a polypeptide of any of claims 1-5.
- A nucleic acid of claim 6, wherein the nucleic acid has a sequence selected from
 the group consisting of nucleic acid sequences represented by SEQ ID NO:3 and SEQ ID NO:6.
 - 8. A method of identifying a small molecule immunosuppressant compound, comprising
- constructing a cell-based two hybrid protein-protein interaction assay, wherein one of two fusion hybrid proteins in a cell contains an (A) subunit of calcineurin, and the other of two fusion hybrid proteins contains a CNI polypeptide,

contacting the cell with a small molecule, and

identifying the small molecule as an immunosuppressant if the molecule potentiates an interaction between the two hybrid proteins.

9. A method of claim 8, wherein the cell is a yeast cell.

5

- 10. A method of claim 8, wherein one of the two fusion hybrid proteins contains a GAL4 activation domain and the other of two fusion hybrid proteins contains a GAL4 binding domain.
- 10 11. A method of claim 8, wherein the subunit of calcineurin is selected from the group consisting of yeast CNA1 and yeast CNA2.
 - 12. A method of claim 8, wherein the subunit of calcineurin is an "A" subunit of human calcineurin.

- 13. A method of claim 8, wherein the CNI polypeptide is yeast CNI polypeptide.
- 14. A method of claim 8, wherein the CNI polypeptide is yeast CNIc polypeptide.
- 20 15. A method of claim 8, wherein the cell is further modified to cause overexpression of a "B" subunit of calcineurin by said cell.
- 16. A yeast cell carrying a mutation in the naturally-occurring genomic copy of a gene encoding calcineurin-interacting polypeptide, where said mutation prevents expression of a
 25 functional calcineurin-interacting polypeptide from said genomic copy.

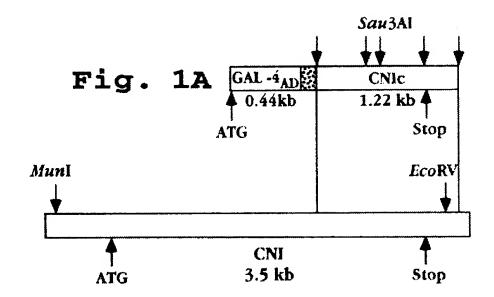
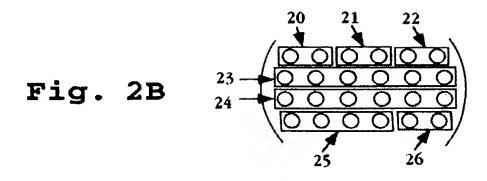


Fig. 1B





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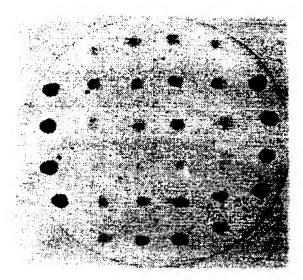


Fig. 3A

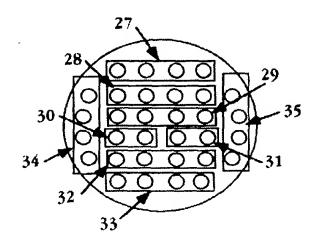
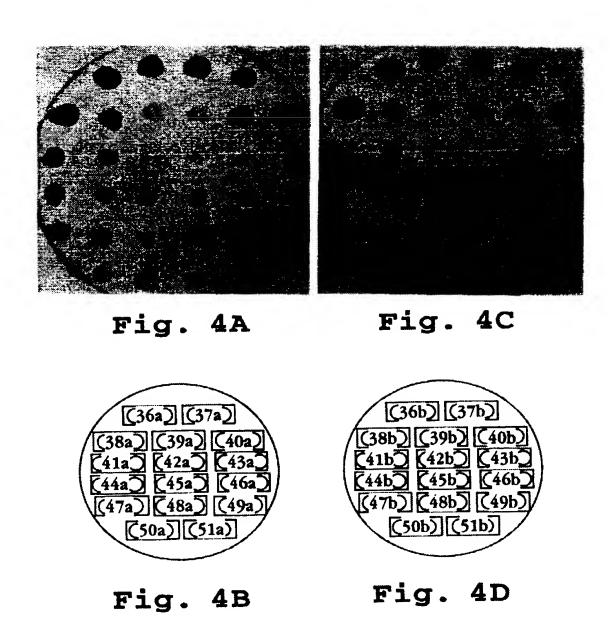
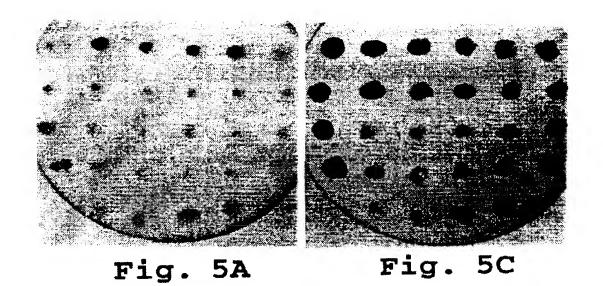
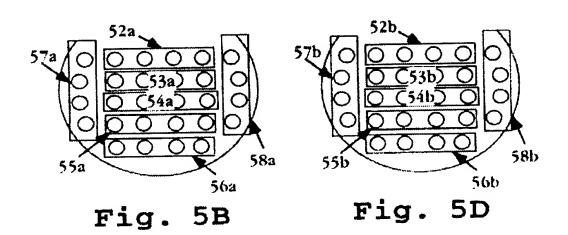
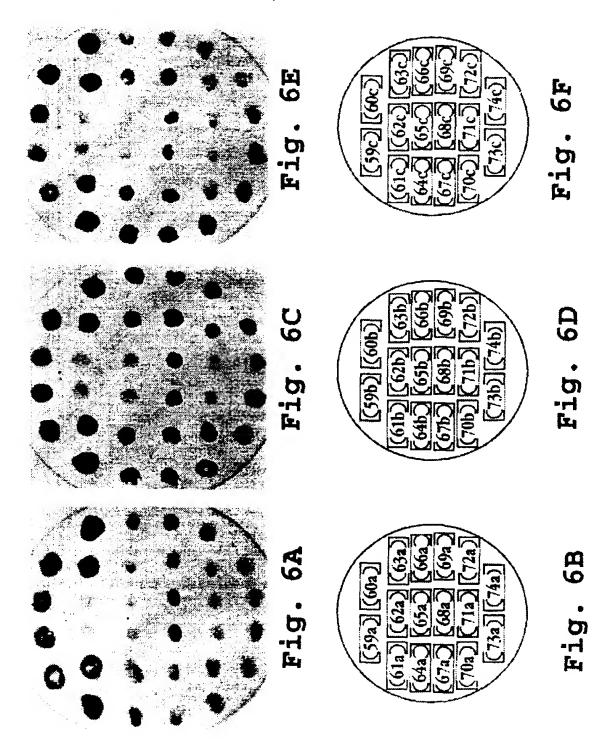


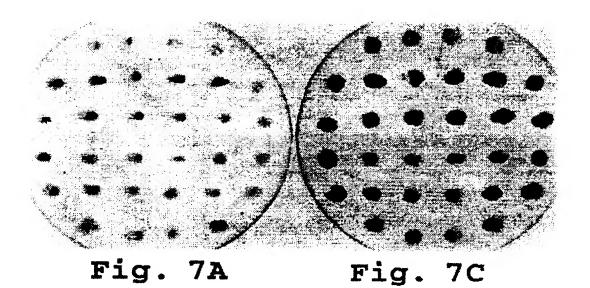
Fig. 3B

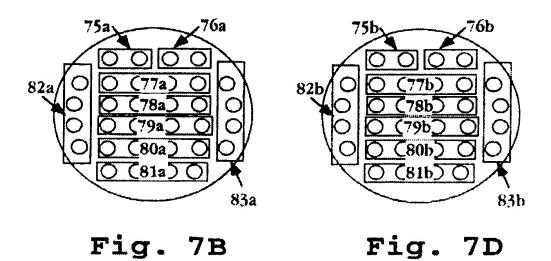












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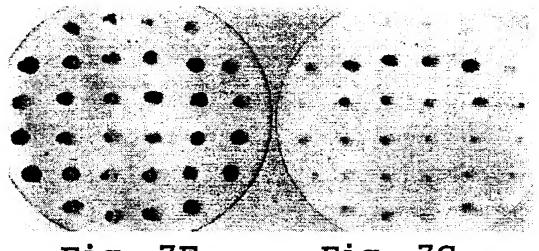


Fig. 7E

Fig. 7G

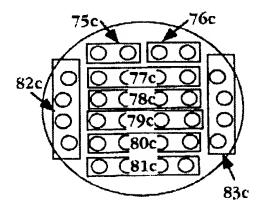


Fig. 7F

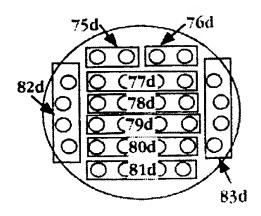


Fig. 7H

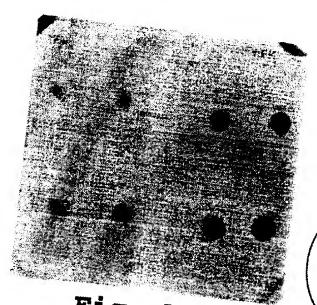


Fig. 8A

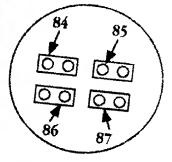


Fig. 8B



Fig. 9A



Fig. 9C

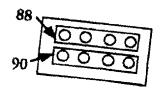


Fig. 9B

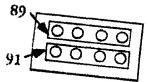
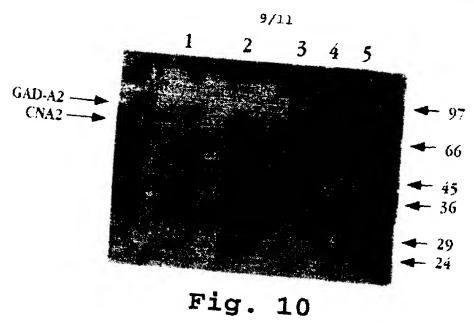


Fig. 9D

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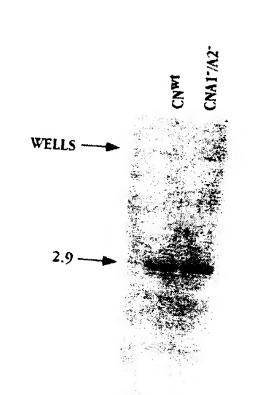


Fig. 11

tttccagcgg catatcgtgg ctttcaccga acacttcctt cgagagagtg cattttacta 61 tgtgaaccaa tttttcctct ttttcggttt gcaagttcac ctgaaaaact gcttaacact 121 actagcaatt gccctattgt cgtacgagga ctttgccaaa tgtattcccg gctgtttgta 181 gtatatatac gcagatatat aatagcgccg tctttttacc tctttgagcg aattgccaaa 241 tattgactet tittgicttat titegetatee ceatettate aaaaatggga acaactegit 301 gaaataagag acaagcaaca agaaagacaa ccaacagaaa gttccattcc gcacaaatac 361 getggaatee catagaatat tgettgttee tetatgaeta catgeteeaa tteaatacag 421 aaaatgatac tgtagctcca gtgtttccca tggagcaaga tataaatgca gcacctgatg 481 cogtoccact ggtgcagaca acaacactac aagtotttgt aaagottgcc gaacccatag 541 tgtttttaaa aggatttgaa actaacggac tgtctgaaat agcccccagt atcttacgag 601 gatetettat egteagggtg ttgaaacega ataaattaaa aagtatateg ataacettea 661 aaggaatatc cagaacagag tggccggaag gtataccacc gaagagagaa gaattttcag 721 atgttgaaac tgttgtcaat cacacatggc cattttatca ggcggatgac ggcatgaatt 781 ctttcacctt agaacatcac agctcaaata attcgtccaa tcgcccatct atgagcgatg 841 aagattatet aettgaaaaa ageggtgett cagtatatat cecaccaace getgaacece 901 ctanagataa tagcaatcta agtctggatg cctatgagcg caactcattg tcatccgata 961 atttgagtaa caagccagta tcaagtgatg tttcccatga cgacagtaaa ctgttggcta 1021 ttcamaagac accattacca tcatctagtc gaagaggatc ggtaccggca aattttcacg 1081 gtaactettt gteaceteat acetteatat etgatttgtt cacaaaaaca tteagtaata 1141 gtggcgctac tccaagtcct gagcaagagg ataactatct tacaccatcc aaagattcta 1201 aagaagtttt tatttttega cegggegatt atatttacae ttttgaacag ceaatatege 1261 aatottatoo agaaagtata aaagocaatt ttggttoogt ggagtataaa ctgtoaatag 1321 acatagagag gtttggcgca ttcaaatcaa ctatacatac tcaattaccc atcaaagtcg 1381 taaggettee ttetgatgga teegtagaag agaetgaage tattgcaatt teeaaggaet 1441 ggaaagatet tetteattat gaegtggtaa tittetegaa agagategit tigaatgeat 1501 tittacccat cgatttccat ttcgctcctc tagataaagt tactctgcat cgtattagaa 1561 tttatctaac agagtctatg gaatacactt gtaatagtaa tggaaatcac gagaaggctc 1621 gtagattaga gccaactaaa aagtttetgt tggetgaaca taaeggteet aaactgeete 1681 atataccago tggttegaat cotttgaagg ctaaaaatag agggaacato ctottggatg 1741 aanaatcegg egatetagtt aacaaagatt tteagttega ggtgtttgte eeaagcaagt 1801 ttacaaacag tatacggtta caccctgata caaattatga taaaatcaaa gcccaccatt 1861 ggataaaaat ttgccttcgt ctttccaaga agtacgggga caatagaaaa catttcgaaa 1921 taagtattga ttctccaatc catattttaa atcaactatg ctcacacgcg aatactttgc 1981 taccgageta cgagagteat ttccagtatt gtgatgaaga tggtaatttc gcaccagcag 2041 cagatcaaca aaattacgca agtcatcatg attccaatat tttcttccca aaagaagttc 2101 tttcgtctcc cgttctttca cctaacgtgc agaagatgaa cattagaata ccgtctgatc 2161 ttccagtagt gegtaataga getgaaageg taaagaaaag caagteagat aataceteea 2221 agaagaatga tcaaagtagc aatgtottog catocaaaca gotggtogca aacatttata 2281 ageceaatea gatteeaaga gaattaaett eteeteagge gttaceatta tegeceatea 2341 ceteaceaat teteaattac caaceattat caaacteece geeteeagat titigattitig 2401 atctagctaa gegeggegea geegattete atgetattee tgtggateet ecateatatt 2461 ttgatgtatt aaaggeegat gggattgaat tgeeatacta egatacaagt teatetaaaa 2521 ttcctgaact aaaactaaac aaatctagag agacattggc cagcattgag gaggactcat 2581 tcaatggttg gtctcaaatt gatgacttat ccgacgaaga tgacaatgat ggcgatatag 2641 catctggttt caacttcaag ctgtcaacca gtgctccgag tgagaacgtt aattcacaca 2701 ctcctatttt gcagtcttta aacatgagtc ttgatgggag aaaaaaaaat cgtgccagtc 2761 tacacgcaac atcagtgtta cctagtacaa taagacagaa caatcagcat ttcaatgaca 2821 taaaccagat gctaggcagt agtgacgaag atgcctttcc caaaagccaa tcattaaatt 2881 tcaataagaa actaccaata cttaaaatta atgataacgt catacaatca aacagcaata 2941 gtaataacag agttgataat ccagaagata cagtggattc ttcagtcgat attacagcat 3001 tttatgatcc aagaatgtca tcagattcca aatttgattg ggaggtaagc aagaaccatg 3061 ttgacccage agectacteg gttaacgttg ctagtgaaaa cegtgtactg gacgacttta 3121 agaaagcatt tegegaaaag agaaaataag tacattattt teatteteeg acagaattge 3181 taccatttta ctttgtgtcc tgtgattcaa tagtgtacaa tatattggac attttatagt 3241 atacaaatat acaccatcaa totatacato catatoactt gtogtaaaga tatocotttt 3301 taatagtaca gcgattaaaa aaataacatg attaacgt

Fig. 12

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RRGSVPANFHGNSLSPHTFISDLFTKTFSNSGATPSPEQEDNYLTPSKDSKEVFIFRPGDYIYTFEQPISQS

YPESIKANFGSVEYKLSIDIERFGAFKSTIHTQLPIKVVRLPSDGSVEETEAIAISKDWKDLLHYDVVIFSK
EIVLNAFLPIDFHFAPLDKVTLHRIRIYLTESMEYTCNSNGNHEKARRLEPTKKFLLAEHNGPKLPHIPAGS

NPLKAKNRGNILLDEKSGDLVNKDFQFEVFVPSKFTNSIRLHPDTNYDKIKAHHWIKICLRLSKKYGDNRKH

FEISIDSPIHILNQLCSHANTLLPSYESHFQYCDEDGNFAPAADQQNYASHHDSNIFFPKEVLSSPVLSPNV

CNIC

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PILNYQPLSNSPPPDFDFDLAKRGAADSHAIPVDPPSYFDVLKADGIELPYYDTSSSKIPELKLNKSRETLA

SIEEDSFNGWSQIDDLSDEDDNDGDIASGFNFKLSTSAPSENVNSHTPILQSLNMSLDGRKKNRASLHATSV

LPSTIRQNNQHFNDINQMLGSSDEDAFPKSQSLNFNKKLPILKINDNVIQSNSNSNNRVDNPEDTVDSSVDI

TAFYDPRMSSDSKFDWEVSKNHVDPAAYSVNVASENRVLDDFKKAFREKRK

Fig. 13

ONAL SEARCH REPORT Application No

PCT/US 95/13580

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B. FIELDS	o International Patent Classification (IPC) or to both national classification (IPC) or to both national classification system followed by classification system followed by classification C12N C07K G01N C12Q		
	tion searched other than minimum documentation to the extent the	it such documents are included in the fi	icids searched
	lata base consulted during the international search (name of data t	sees and where practical search terms	used)
Electronic d	lata base consulted during the international search (name of consu		
C. DOCUM	MENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the	relevant passages	Relevant to claim No.
x	Swissprot Database entry YK01_Y Accession number P36117; June 0 Duesterhoeft A. et al.:'Hypothe kD protein in YPT52-GCN3 interg region.'	1,1994 tical 102.5	1-5
X	Emfun Database entry Scykr021w Accession number Z28246; May 10 Duesterhoeft A. et al.:'S. cere chromosome XI reading frame ORF	visiae	6,7
P,X	MOLECULAR BIOLOGY OF THE CELL, 1994. 141A., HUANG L ET AL 'A novel protein interacts with calcineurin in v see abstract 818	that	1,2,6, 8-15
X Fur	ther documents are listed in the continuation of box C.	Patent family members are	listed in annex.
'A' documents 'E' earlier filing 'L' documents 'C' documents 'O' documents 'P' documents	nent which may throw doubts on priority claim(s) or it is cited to establish the publication date of another on or other special reason (as specified) ment referring to an oral disclosure, use, exhibition or means the prior to the internanonal filing date but	"T" later document published after or priority date and not in concided to understand the princip invention." 'X' document of particular relevant cannot be considered novel or involve an inventive step when cannot be considered to involve document is considered to involve document is combined with on ments, such combination being in the art. '&' document member of the same	ce; the claimed invention cannot be commerced to the claimed invention cannot be commerced to the document is taken alone or; the claimed invention e an inventive step when the se or more other such docugate or so a person skilled
Date of the	than the priority date claimed e actual completion of the international search 21 March 1996	Date of mailing of the internation of 2. 04.	coal search report
	mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijk Tel. (+ 31-70) 340-2040, Tx. 31 651 epo ml, Fax: (+ 31-70) 340-3016	Authorized officer Espen, J	

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rmational Application No PCT/US 95/13580

(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT ategory * Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.	
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